

1 Temperature diagnostic to identify high risk areas and optimize *Legionella pneumophila*
2 surveillance in hot water distribution systems

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28 **Abstract**

29 *Legionella pneumophila* is frequently detected in hot water distribution systems and
30 thermal control is a common measure implemented by healthcare facilities. A risk
31 assessment based on water temperature profiling and temperature distribution within
32 the network is proposed, to guide effective monitoring strategies and allow the
33 identification of high risk areas. Temperature and heat loss at control points (water
34 heater, recirculation, representative points-of-use) were monitored in various sections
35 of five healthcare facilities hot water distribution systems and results used to develop a
36 temperature-based risk assessment tool. Detailed investigations show that defective
37 return valves in faucets can cause widespread temperature losses because of hot and
38 cold water mixing. Systems in which water temperature coming out of the water
39 heaters was kept consistently above 60°C and maintained above 55°C across the
40 network were negative for *Legionella* by culture or qPCR. For systems not meeting these
41 temperature criteria, risk areas for *L. pneumophila* were identified using temperature
42 profiling and system's characterization; higher risk was confirmed by more frequent
43 microbiological detection by culture and qPCR. Results confirmed that maintaining
44 sufficiently high temperatures within hot water distribution systems suppressed *L.*
45 *pneumophila* culturability. However, the risk remains as shown by the persistence of *L.*
46 *pneumophila* by qPCR.

47

48 **Key words**

49 *Legionella pneumophila*, premise plumbing, viable but not culturable (VBNC), heat
50 treatment, temperature profile, culturability.

51

52 **Highlights**

- 53
- 54 • Temperature profiles were generated for hot water distribution systems points-
55 of-use
 - 56 • Risk assessment based on temperature profile results at control points was
57 developed
 - 58 • *L. pneumophila* positive areas were predicted using the risk assessment tool
 - 59 • A temperature diagnostic flowchart is proposed to identify *L. pneumophila* risk
60 areas

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65 **1. INTRODUCTION**

66 *Legionella pneumophila* (*Lp*) is an opportunistic pathogen that can proliferate in hot
67 water distribution systems (HWDS) of large buildings, such as health care facilities
68 (HCFs), where it can cause waterborne nosocomial pneumonias. Although its optimal
69 growth temperature lies between 25 and 42°C (Yee and Wadowsky 1982), *Lp* has been
70 isolated from water systems at temperatures up to 60°C (Martinelli et al. 2000), and in
71 cold water systems with temperatures below 20°C (Arvand et al. 2011). The presence of
72 *Lp* in HCFs water systems is well demonstrated, with reports of 10 to 50% positive hot
73 water samples taken from taps and showers in Europe and the United States (Arvand et
74 al. 2011, Bargellini et al. 2011, Martinelli et al. 2000, Serrano-Suarez et al. 2013, Stout et
75 al. 2007). Risk characterization of water sources remains uncertain because of the lack
76 of reliable dose-response models (Buse et al. 2012) and therefore the difficulty to define
77 an acceptable level of *Lp* contamination that would minimize risk. While the level of
78 positivity for *Legionella* in health care facilities (HCF) HWDS has been proposed as a
79 reliable predictive risk factor (Best et al. 1983, Lin et al. 2011), the specificity and
80 sensitivity of the 30% positivity cut-off point has been recently questioned (Allen et al.
81 2012, Allen et al. 2014, Pierre et al. 2014).

82

83 Control of *Legionella* risks in health care facilities (HCFs) is addressed and regulated
84 through guidance documents (Bartram et al. 2007, BSR/ASHRAE 2013, CDC 2003, HSE
85 2013, République Française 2010a). System characterization and environmental
86 monitoring are among the first steps to establish a water safety plan or to evaluate the

87 operational risk in hot water distribution systems (HWDSs), especially in HCFs
88 (BSR/ASHRAE 2013, Department of Health (DH) and Estates and Facilities Division 2006,
89 République Française 2010b, WHO 2011). Recent guidelines stress the need to properly
90 manage hydraulics to ensure homogeneous temperature and biocidal control in all
91 areas of the HWDS (CSTB 2012), and system balancing under varying demand should be
92 verified.

93

94 Although a multitude of possible system architectures are encountered, a simplified
95 schematic of a hot water distribution system (HWDS) can be established (Fig. 1) and
96 should include: the number and characteristics of key systems components such as the
97 calorifiers, reheating units and reservoirs, the distribution systems including principal,
98 subordinate and tertiary flow and return loops and point-of-use devices (tertiary
99 terminal end). A schematic and characterization of each HWDS within a premise must
100 be established independently (BSR/ASHRAE 2013, HSE 2013, République Française
101 2010a). This data is the foundation for interpreting monitoring results and identifying
102 high risk areas.

103

104 A summary of the key elements from selected regulations and guidelines to implement
105 temperatures control of L_p in large buildings, and when available, in HCFs is provided as
106 supplementary material (Table S1). Approaches to control L_p in hot water distribution
107 systems (HWDSs) vary considerably, but all guides include objectives or obligations for
108 optimal operating temperatures at critical points in the distribution systems. Also

109 commonly specified are construction and operational standards, such as minimizing
110 stagnation (recirculation loops, elimination of hydraulic and physical dead ends, etc.),
111 recommendations on the use of devices and materials not promoting bacterial
112 proliferation (construction material, flow, temperature, etc.) and requirements for
113 microbiological monitoring in relation to pre-established criteria that define corrective
114 actions.

115

116 In France, recently strengthened regulations determine mandatory minimum
117 temperature and *Legionella* monitoring at defined critical control points: 1) hot water
118 outlet and reservoir when present; 2) return loop; and 3) representative points-of-use
119 considered at risk (farthest from the water heater or serving vulnerable patients) but
120 the number of sampling points to be monitored is not specified (République Française
121 2005, 2010a, b, Table S1). It is recommended that temperatures be monitored daily or
122 continuously at hot water heater outlets and at each return loops, and weekly at service
123 points in HCFs. Temperature measurements at points of use are conducted on flushed
124 samples (2-3 min). In the United Kingdom, a risk management approach is proposed,
125 with recommended preventive measures including system maintenance, elimination of
126 stagnation or dead zones, reduction of aerosol formation, maintenance of adequate
127 temperatures and use of materials unfavorable to biofilm development (Department of
128 Health (DH) and Estates and Facilities Division 2006, HSE 2013). Temperature control
129 regimen is presented as the preferred initial approach for *Legionella* control (Table S1).
130 Minimal monthly temperature monitoring is specified at control points including water

131 heater outlet, return loops and sentinel taps. Sentinel taps include representative at-
132 risk taps as well as the first and last taps of each return loops. The use of continuous
133 temperature monitoring is recommended for the water heater outlets and the return
134 loops. In addition, temperature at the tap should be monitored annually on a rotating
135 basis covering 20% of taps yearly, to ensure the whole system is meeting required
136 temperatures for *Legionella* control. It is not permissible to shut down pumped
137 recirculation as it would lead to the loss of the required system temperatures.
138 *Legionella* monitoring is not prescribed unless target temperatures cannot be achieved;
139 however it is recommended in areas with highly vulnerable patients. Weekly flushing for
140 several minutes is recommended for low usage taps.

141

142 Although all available regulations and guidelines provide information on various aspects
143 of the implementation of a successful temperature control regimen, there is no
144 consistent guidance on key elements such as the selection of sentinel points, the
145 incorporation of *Lp* monitoring and the interpretation of the temperature monitoring
146 results. Reports on the efficacy of the implementation of temperature control in health
147 care facilities (HCFs) reveal limited success (Arvand et al. 2011, Bargellini et al. 2011,
148 Blanc et al. 2005, Darelid et al. 2002, Hrubá 2009, Lee et al. 2011, Serrano-Suarez et al.
149 2013). Nevertheless, adjusting the temperature at the heater outlet to ensure water
150 temperatures greater than 50-55°C at distal outlets can be highly effective in reducing
151 the proportion of positive swabs or water samples (Arvand et al. 2011, Blanc et al. 2005,
152 Ezzeddine et al. 1989). Moreover, areas consistently positive for *Lp* were associated with

153 poor hot water recirculation leading to temperature losses (Blanc et al. 2005). In most
154 case studies, the actual conditions of application of the temperature control regimen
155 are poorly documented with some information on temperatures only available for the
156 water heater and return. The efficacy of temperature control regimens must be
157 assessed by its ability to suppress *Lp* growth in the distal areas, as distal growth is highly
158 significant (Cristina et al. 2014, Serrano-Suarez et al. 2013). On the other hand, there is
159 increased risk of scalding for temperatures higher than 50°C at the tap (Moritz and
160 Henriques 1947). Some countries specify maximum temperatures at the point-of-use to
161 avoid scalding (Table S1), but newly updated regulation in United Kingdom require a risk
162 assessment comparison between the risk of scalding and the risk of infection before
163 limiting the hot water temperature below 50°C, a risk factor for *Legionella* proliferation.

164

165 Although the critical elements of temperature control in guidelines and regulations to
166 reduce *Legionella* risks in HWDSs rely on scientific evidence and application experience,
167 the detailed implementation, especially the selection of critical control points and
168 monitoring requirements, most often reflect economic constraints. In addition,
169 significant discrepancies exist between proposed modalities of implementation and
170 management. The objectives of the present study were to: (1) demonstrate the
171 potential of detailed temperature profiling to identify areas at risk of *Lp* in the hot water
172 distribution systems (HWDSs) of five health care facilities (HCFs); (2) identify effective
173 monitoring strategies and guidance to conduct temperature profiling and interpret

174 monitoring results; (3) propose a risk characterisation approach based on temperature
175 diagnostic at critical control points.

176 **2. MATERIALS AND METHODS**

177 **2.1. Hot water system characterization**

178 Five hot water distribution systems (HWDSs) were analyzed. Systems 1 to 4 are smaller
179 systems within a 7-story general hospital facility of 255 beds using conventional electric
180 water heater being fed chlorinated ground water. System 5 has a larger flash system
181 feeding a ten-story 450 bed children's hospital fed by surface filtered chlorinated water.
182 A survey of the different HWDSs and connected units was first completed.

183

184 The principal flow and return loop of each system was sampled at the water heater
185 outlet, in the principal return loop and prior to the return point into the water heater.
186 The sampling ports were seldom used and were flushed prior to sampling to ensure no
187 stagnant water from the sampling port would be collected. The sampling port was
188 cleaned with ethanol and sterilized MilliQ water. Two samples were collected at each
189 point: 1) 2L in sterile polypropylene bottles with sodium thiosulfate (final concentration
190 of 1.1mg/L) microbiological analysis and 2) 250 mL for pH, temperature, chlorine and
191 conductivity measurements. Municipal water feeding the hot water systems was
192 sampled following the same protocol. In addition continuous temperature monitoring
193 was conducted on 3 subordinate return loops for system 5, using a Datalogger (RDXL4SD
194 4-Channel, Omega, Qc, Canada).

195

196 **2.2. Temperature profiling and water sampling at points-of-use**

197 Sentinel taps where sampling was performed were selected based on the following
198 criteria: representative of different building levels, some at the far end and preferably in
199 areas serving vulnerable patients such as intensive care units, surgical ward, transplant,
200 infectious diseases. All sampling events were conducted between July 2012 and October
201 2013. The first part of the sampling campaign was conducted to establish temperature
202 profiles at each selected sentinel point-of-use in hot water, across all 5 systems between
203 July 2012 and March 2013. A temperature probe was inserted into the water to
204 measure the temperature over a 20 minute period of continuous flow. Each system had
205 a number of taps sampled proportional to the size of the system. Systems 1 to 5 had
206 respectively 3, 6, 3, 7 and 36 taps sampled. The temperature profiles could not be
207 generated for 1 mitigated tap in system 1, 3 in system 2 and 1 in system 3. In addition,
208 three taps from system 5 were selected for a repeat temperature profile sampling.
209 Residual chlorine was measured onsite (Pocket Colorimeter™ II, Hach, USA) for all
210 samples.

211

212 The second part of the sampling campaign was conducted to evaluate the presence of
213 *Lp* at the point-of-use. All sentinel points of systems 1 to 4 and 8 sentinel points from
214 system 5 were sampled for microbiological analysis. Sentinel points from system 5 were
215 selected based on temperature profile results. For each sampling point, 3L of hot water
216 were collected without prior flush into sterile polypropylene bottles containing sodium

217 thiosulfate (final concentration of 1.1 mg/L). Of the 3 liters collected, 1L was used for
218 culture, 1L for qPCR and 1L was collected as extra. This sampling was repeated 4 times
219 at 3 selected sampling points in 2 systems fed by the same source water: a system with
220 no positive sites for *Lp* (system 1) and a system with a high positivity rate (system 4).
221 The 3 control points selected were the water heater outlet, one representative tap and
222 the principal return loop.

223

224 **2.3. Impact of stagnation**

225 The third part of the sampling campaign was conducted on 2 taps of system 5 to study
226 the impact of stagnation on the detection of *Lp*. Stagnation is defined as the period
227 during which the tap is not used and water remains idle within the piping. One liter of
228 hot water was sampled in sterile propylene bottles with sodium thiosulfate (final
229 concentration 1.1 mg/L) after 1 hour, 1, 2, 3, 5 & 10 days of stagnation. These
230 stagnation times were chosen to represent various situations within a real system: 1h
231 for the time between usage in a patient room; 1-day for patient daily care; 2 and 3 days
232 for areas closed on the weekend (i.e. outpatients clinics); 5 and 10 days for an empty
233 room in between patients or a temporary ward closure. *Lp* concentrations were
234 measured by qPCR as described in section 2.4.

235

236 Heat losses during stagnation periods were evaluated in the laboratory, on 81 cm of
237 1.25 cm diameter copper pipes at room temperature (20°C) without insulation and with
238 insulation: Type 1, 2.54 cm thick fiberglass insulation with PVC jacket (Caltech Isolation,

239 Canada) and Type 2, 0.95 cm thick polyethylene foam insulation (Tundra, Industrial
240 Thermo Polymers Limited, Canada).

241

242 **2.4. Microbiological analyses**

243 Water samples were mixed thoroughly and divided to perform isolation and
244 quantification of *Legionella* spp. and *Lp* by culture and quantitative polymerase chain
245 reaction (qPCR).

246

247 Culture was conducted according to the standard AFNOR NF T90-431 procedure (AFNOR
248 2006). Briefly, 1 liter was filtered on sterile 0.4 µm polycarbonate membranes (47 mm
249 diameter; Maine Manufacturing, LLC), which were then sonicated in 5 ml sterile water
250 at 47 kHz for 1 min (Bransonic, Danbury, USA). Heat treatment (50°C, 30 min), acid
251 treatment (pH=2; 5 min) and combination of both were performed on 3 separate 1 ml
252 aliquots. Samples were plated on GVPC selective agar (Innovation Diagnostics Inc.) and
253 incubated at 36°C for 10 days. Typical colonies that developed after 4 to 10 days were
254 sub cultured on confirmation plates for 2 to 4 days at 36°C. Resulting colonies that
255 developed on BCYE agar, but neither on blood agar nor on BCYE without cysteine were
256 considered as *Legionella* spp. Confirmation for *Lp* was conducted using the *Legionella*
257 latex test (DR0800, OXOID Limited). The calculated detection limit for the culture
258 method was 50 CFU/L for both *Legionella* spp. and *L. pneumophila*.

259

260 Quantification by qPCR was performed on a Corbett Rotorgene 6000 using the iQ-Check
261 Quanti *L. pneumophila* kit (Bio-Rad, Mississauga, Canada) with the following protocol:
262 15 min initial denaturation at 95°C followed by 50 cycles with denaturation at 95°C for
263 15 s, annealing at 57°C for 30 s, elongation at 72°C for 30s and final elongation for 15
264 min at 72°C (Bonetta et al. 2010). An internal control and four DNA standards ranging
265 between 19 and 3.9×10^4 genomic units (GU) were supplied with the kit. Sterilized water
266 was used as negative control. DNA extraction was performed directly on filters using a
267 bead beating method adapted from Yu and Mohn (1999). Briefly, 1L was filtered on 0.45
268 μm mixed cellulose ester and the filter was inserted into an extraction tube containing a
269 garnet matrix and one 1/4-inch ceramic sphere (Lysing Matrix A, MP Biomedicals, Solon,
270 USA). Lysing buffer was added to each tube prior to the bead beating step performed on
271 a FastPrep MPBio-24, followed by ammonium acetate precipitation and successive
272 ethanol washes.

273

274 **2.5. Statistical analysis**

275 Statistical analyses were performed with Statistica10 (StatSoft). A one-way analysis of
276 variance (one-way ANOVA) was used to evaluate differences between the 3 control
277 points sampled in systems 1 and 4 during the repeat sampling. A t-test was used to
278 detect differences between the two taps sampled at various stagnation times in system
279 5. Significance level was set at $p = 0.05$.

280 **3. RESULTS & DISCUSSION**

281 **3.1. General system characterization**

282 Systems 1, 3, 4 & 5 presented a multiple vertical subordinate flow and return loop
283 configuration feeding in average three devices per story. System 2 was a simplified
284 horizontal architecture with only few vertical pipes feeding water to horizontal
285 subordinate flow and return loops (Fig. 1). There is no reported evidence showing that
286 the vertical or horizontal configuration is a determining factor for the risk of
287 contamination. Other factors including hot water temperature, effective recirculation in
288 the subordinate loop, the presence of dead-ends, piping material and water velocity
289 have been identified as risk factors (Health and Safety Executive (HSE) 2013).
290 Nevertheless, it is important to know and document the configuration of a studied
291 system to interpret temperature data collected. With information on the pipe diameter
292 and configuration, the location and relative importance of recirculating and stagnating
293 volumes can be determined providing information to guide monitoring and control
294 strategies. For example, the recirculated volume was approximately 900L, of which
295 600L in the principal flow and return loop (50 mm diameter) and 300L in the
296 subordinate flow and return loops (10 vertical risers of 25 mm mean diameter). The
297 distal volume in the tertiary terminal end was about 300L (Fig. 1). However, this volume
298 can be minimized if a tertiary return loop is added, leaving only the small connecting
299 volume of less than 150mL per device accounting for a total of 90L of stagnant volume
300 (Fig. 1).

301

302 For systems 1 to 4, incoming water had chlorine residual of 0.30 ± 0.03 mg Cl_2/L , pH of
303 7.77 ± 0.05 and conductivity of 307 ± 29 μS . For system 5, residual chlorine was higher, at
304 0.5 ± 0.1 mg Cl_2/L , pH of 7.82 ± 0.07 and conductivity of 288 ± 13 μS . There was no
305 additional disinfection treatment in any of the hot water systems studied and mean
306 residual chlorine was 0.04 ± 0.02 mg Cl_2/L for all systems.

307

308 **3.2. Temperature monitoring**

309 3.2.1. Water heater outlet

310 Most guidelines specify that target temperatures must be maintained at all times, but
311 seldom do they specify the monitoring requirements of measurement frequency.
312 Periodic temperature readings, even daily measurements, do not provide insurance of
313 temperature maintenance in the hot water distribution system (HWDS), unless the
314 stability of the system's performance has been fully established. Systems seemingly
315 providing water above 60°C based on daily measurements can actually produce lower
316 temperature water for extended periods of time. In fact, the mean temperatures at the
317 water heater outlet for four of the five systems studied were above 60°C , but online
318 temperature monitoring revealed that production temperature was repeatedly below
319 60°C and reached down to 43°C in some cases (Table 1 and Fig. 2). System 1
320 consistently produced water above 60°C while systems 2, 3 and 5 regularly produced
321 water below 60°C at certain periods of the day (Fig. 2). For system 3, temperature was
322 monitored weekly by the operators on Saturday mornings during low water demand

323 providing an average of 62.5°C over a period of 24 months (Table 1). Nonetheless, when
324 online monitoring was performed during a typical weekday, mean temperature was
325 lower (57.8°C). It is also interesting to point out that even a very recently installed
326 system (2011) equipped with a flash heating unit was also subject to periodic
327 temperature drops (System 5, Fig. 2). These observations demonstrate the need to use
328 online monitoring to assess the temperature compliance of a HWDS compared to
329 periodic manual readings of temperature. Daily variations in hot water demand in large
330 HCFs with typical peak flow factors of > 6 (Bujak 2010) can influence the temperature at
331 the water heater outlet depending on the system's capacity. The extent and duration of
332 the non-compliance of the hot water outlet temperature set point is important to
333 consider and has been limited to the sporadic short duration (minutes) events in the
334 German technical rules (Table S1).

335

336 3.2.2. Return loops

337 The return loop at the point closest to the water heater is designated as the furthest
338 point from the water heater and continuous temperature monitoring is often
339 recommended (Fig. 1). It is considered as an indicator of the system's capacity to
340 maintain temperatures throughout the hot water distribution system (HWDS). In the
341 five systems studied, the principal return loop temperatures ranged between 50.4 and
342 58.9°C with varying levels of blending from multiple return loops occurring upstream of
343 the principal return control point (Table 1). Continuous monitoring for 2 months at the
344 return loop manifold for combined returns of units 3&5 (45.7°C), units 1&2 (48.0°C),

345 single return for the kitchen (58.1°C) and for unit 3 prior to merging with unit 5 (46.6°C)
346 revealed wide differences compared to the overall combined return loop (53.9°C).
347 Although a regulated control location (Table S1), temperature at the principal return
348 loop is not indicative of the conditions in all subordinate loops within a complex HWDS if
349 the system is not balanced for all water demand conditions. In such cases, it merely
350 represents the mean temperature of the blended recirculated hot water from various
351 sectors of the HWDS. More specifically, it does not provide any information on the
352 actual levels of recirculation and temperature losses in the various sectors of the HWDS
353 and does not in any way confirm efficient recirculation in all subordinate loops. These
354 results suggest the temperature monitoring of subordinate return loops together with
355 the principal return loop as a tool to identify imbalances within a system and as an
356 ongoing system validation measure.

357

358 Heat losses between the water heater outlet and a remote point will occur during
359 stagnation (if recirculation is not effective or shut down for energy conservation
360 purposes) or during circulation in the principal and subordinate flow and return loops.
361 During low demand conditions, recirculation will dictate residence time and drive heat
362 losses. Mean system heat losses were evaluated for each of the five studied systems
363 (Table 1). For three of the five systems, temperature losses between the water heater
364 and the principal return loop mean temperatures exceeded the target of $\leq 5^{\circ}\text{C}$ set in
365 several guidelines (Table S1). Heat losses during circulation can be minimized by
366 reducing residence time. Water velocity can be set to meet desired maximum heat

367 losses and general recommendations suggest maintaining a minimal velocity of 0.2 m/s
368 (Blokker et al. 2010, CSTB 2012), which would result in approximately 30 min residence
369 time and 5°C heat losses in large health care facilities (HCF) insulated HWDS. Although
370 insulation minimizes heat losses under flowing conditions, it is not sufficient to maintain
371 high temperatures over prolonged periods of stagnation. Actually, slower heat losses
372 during stagnation may lead to sustained optimal temperatures for *L. pneumophila*
373 growth. Figure 3 shows that temperature decreased from 60°C to below 50°C within 30
374 min in fully insulated copper pipes and within 10 min for non-insulated pipe, both
375 reaching room temperature after 3.5 hours. Periods of stagnation of 30 min or more are
376 expected in the connecting piping upflow of points-of-use and in areas of inefficient
377 recirculation.

378

379 Existing standards and guidelines set design and operational obligations to control heat
380 losses in hot water distribution systems (HWDS) to maintain at minimum target
381 temperatures throughout the HWDS and to meet energy conservation goals, but these
382 are generally only compulsory for new buildings. Recirculation flow rates should be
383 calculated to maintain a <5°C system heat loss or to ensure a minimum temperature of
384 50-55°C at the end of the return loop assuming adequate recirculation throughout the
385 system (ASPE 2008). The control points results required to evaluate heat loss goals
386 compliance include the principal and subordinate return loops, the most distant point of
387 the flow loop or all points of the system (Table S1). Monitoring results from the five
388 HWDSs clearly show that the selection of the return loop reference point is critical. Heat

389 loss evaluation from the principal return loop may mask major heat losses in
390 subordinate flow and return loops, as we observed in system 5 with losses ranging from
391 3.5 to 16.3°C when evaluated for single or dual subordinate return loops (Table 1).
392 Indeed, wide differences in temperature can occur between secondary return loops,
393 and thus all return loops should be considered individually. The overwhelming
394 importance given to temperature maintenance has also led to the specific banning of
395 recirculation shutdown in Austria and United Kingdom (Table S1). The nightly shutdown
396 of recirculation for energy conservation purposes is only allowed in two rules
397 (CMMQ/RBQ 2013, DVGW German Technical and Scientific Association for Gas and
398 Water 2004) and only with the demonstration of unobjectionable hygienic conditions.
399 Our results point out that the temperature losses of isolated subordinate loops during
400 stagnation resulting from such shutdowns would quickly generate durable temperature
401 conditions favorable to the growth of *Lp*. More importantly, such shutdowns during low
402 or nil demand conditions expose the whole HWDS, instead of a relatively small volume
403 (1,200L versus 90 to 300L in System 5) to these undesirable temperature conditions.

404

405 3.2.3. Temperature distribution at point-of-use

406 Sequential volume profiling results identify in which sections of the HWDS the heat
407 losses take place, namely the tap and its connecting piping, the secondary piping, the
408 distribution columns and/or the main feeder pipes. Profile variability for a given
409 sampling point at different times and days was found to be small, with overall profile
410 and maximum temperature reached being consistent over time despite variable

411 temperature in the first liter (Fig S1). Temperature profiles obtained on the studied
412 systems are summarized in three groups (Fig. 4), with detailed profiles presented in Fig.
413 S2. Systems 1, 2 and 3 (Fig. 4a) met recommendations for water heater outlet and
414 return loop temperatures, with 86% of points reaching 55°C and all points being above
415 50°C after 2 minutes of flow, indicative of limited stagnant water volumes and effective
416 recirculation. Ideal systems should have no or very little transition and reach equilibrium
417 at recommended temperatures in order to maintain sufficient temperatures within the
418 whole system. Despite reaching equilibrium temperature rapidly (<60s), system 4 could
419 not achieve recommended temperature at the points-of-use with 57% of points never
420 reaching 55°C although all above 50°C, mainly due to the insufficient water temperature
421 at the water heater outlet (Fig. 4b). System 5 shows a longer transition period before
422 reaching temperature equilibrium and is unable to meet 55°C for 47% and 50°C for 19%
423 of points, despite water heater and principal return loop temperatures meeting
424 recommendations (Fig. 4c).

425

426 Additional temperature monitoring using surface thermocouples on subordinate flow
427 and return pipes were conducted on system 5 (Fig. S3). The ongoing temperature
428 monitoring in subordinate flow and return loops in addition to the principal flow and
429 return loop provided helpful information to identify local issues. For instance, broken
430 valves in a shower faucet resulted in cold water entering the hot water feed pipe and
431 riser. Fixing the device increased the minimal temperature by an average 5°C in all 10
432 subordinate risers in this wing (Fig. S3, a-c). A second example was insufficient

433 recirculation causing a significant heat loss during night flow, which was corrected by
434 the addition of a local pump on the subordinate return loop, after the furthest pair of
435 risers (Fig. S3, d-g). These examples show the importance of characterizing local
436 conditions and the potential of single faulty devices to influence temperature
437 maintenance in large sections of hot water distribution systems (HWDS). Again, we
438 conclude that relying on temperature maintenance in the principal return loop is not
439 sufficient to identify such risk areas.

440

441 **3.3. *Legionella* monitoring**

442 Results of microbiological measurements for the five studied systems are presented in
443 Table 1. Detection by qPCR was used in complement of culture detection as it has been
444 shown to be efficient in monitoring changes in the bacterial numbers (Krojgaard et al.
445 2011, Lee et al. 2011). Culture positive samples for *Lp* were detected in systems 4 and 5
446 with 22 and 27% positivity respectively (detection limit = 50 CFU/L; quantification limit =
447 250 CFU/L). Culture positive samples results were low, with only one count above
448 quantification limit at 600 CFU/L, located at a tap in system 5. Positivity increased above
449 80% for both systems when measured by qPCR and remained below detection limit for
450 systems 1-3, except for one sample in system 2 (Table 1, Table S2). Systems in which
451 water temperature was kept consistently above 60°C coming out of the water heater
452 and maintained above 55°C across the network were below detection limit for
453 *Legionella* by culture or qPCR. Such results strongly suggest that satisfactory
454 management of temperature at control points in the studied systems resulted in lower

455 prevalence. However, these results represent a water quality snapshot at a point in time
456 and are not necessarily representative of microbial quality over time or at other
457 locations in the HWDS. Several factors affecting *Lp* densities at a given point have been
458 identified including intrinsic biological system heterogeneity, culturability, prior
459 stagnation and sample volume. Napoli et al. showed variation of $\leq 20\%$ concentrations
460 of CFU/ml from one day to the next within a ward during repeated sampling over five
461 consecutive days across eight units within a hospital (Napoli et al. 2009). In the present
462 study, confirmation sampling was conducted in two of the five HWDSs to investigate the
463 temporal variability. Fig. 5a shows results from repeated sampling conducted at three
464 control points (water heater outlet, principal return loop and a point-of-use) in systems
465 3 & 4. All samples were negative in qPCR and culture for system 1, whereas samples
466 from system 4 were consistently positive in qPCR and to a lesser degree in culture (Fig.
467 5a). Mean levels of *Lp* detected in system 4 were not significantly different between the
468 3 control points ($p > 0.05$). These findings are in agreement with recent reports of
469 discrepancies between trends in *Lp* by qPCR and culture in suboptimal conditions for
470 inactivation of viable but not culturable (VBNC) cells (Krojgaard et al. 2011, Lee et al.
471 2011). Krojgaard et al. showed that qPCR levels can be used to verify the impact of
472 corrective actions such as thermal shock and demonstrated non-detects qPCR results as
473 a predictor of low risk.

474

475 Another factor that may influence levels of *Legionella* in water is the duration of
476 stagnation prior to sampling. Recent evidence reported an increase in bacterial

477 concentrations after various stagnation times (overnight to 14 days) in the cold water
478 distribution system of a large building (Lautenschlager et al. 2010, Lipphaus et al. 2014).
479 A steady increase was observed in the first 12 hours of stagnation whereas longer
480 stagnation time did not lead to further increase (Lautenschlager et al. 2010). In the
481 present study, hot water was sampled from two taps at different stagnation times and
482 *Lp* concentration was evaluated by qPCR (Fig. 5b). The taps were not found to be
483 statistically different when comparing mean results and no correlation was established
484 between the mean *Lp* concentration and the stagnation time. However, the stagnation
485 times were longer than 12 hours, except for the 1h stagnation and samples were taken
486 from the hot water systems. To our knowledge, there is no reported data on the impact
487 of stagnation on bacterial concentrations in hot water. These results suggest that *Lp*
488 concentrations in the first liter of hot water at the tap may not be affected by stagnation
489 time.

490

491 The volume of sample determines the source of the water within the HWDS. *Lp*
492 monitoring can be performed to assess the risk associated with 1) the water heater and
493 primary distribution network using flushed samples, and 2) the distal system, including
494 the tap and its connection to the main distribution system, using samples collected
495 without prior flushing. Cristina et al. (2014) reported that distal stagnation increased the
496 number of positive sites from 2.63 % to 15.79% and mean concentration from 7 vs 637
497 vs CFU/L for *Lp* sg1. Such distal amplification was not as clearly observed by these
498 authors for *Lp* sg2-14 with 40.79 % to 42.11% positive and mean concentration from

499 19,455 vs 26,746 CFU/L. Similar trends were observed for *Legionella* spp in HWDS taps
500 with increased concentration from 45 CFU/L (23% positivity) after a 3 minute flush to
501 226 CFU/L (35% positivity) in the first liter (Serrano-Suarez et al. 2013).

502

503 Although post-flush samples provide insight into systemic hot water distribution system
504 (HWDS) contamination, results from the first volume to flow are indicative of the acute
505 concentrations to which patients may be more readily exposed. In the first volume to
506 flow from the tap, water temperature will often be lower due to previous stagnation
507 and disinfectant will be absent, favoring culturability of cells. On the other hand, higher
508 copper concentration present after prolonged stagnation could impact culturability.
509 Non-detection of *Lp* by culture at a given sampling point and time doesn't necessarily
510 equate to absence of risk for the system.

511

512 Volume sampled, typically 1L or more for *Lp*, plays an important role in data
513 interpretation, either for temperature measurements or microbiological detection
514 where the detection limit of the method improves with the use of higher volume of
515 samples. As illustrated on Fig. 1, sampling the first liter will collect water from the tap
516 and connecting pipes, and might reach water from the subordinate return and flow
517 pipes depending on the configuration. For example, 8 meters of a 13 mm diameter pipe
518 are required to reach 1L. If a larger sample volume is required to do multiple analyses
519 (i.e. culture and qPCR or simultaneous detection of other waterborne opportunistic

520 pathogens), it should be kept in mind that water will become less representative of the
521 point-of-use.

522

523 **3.4. Value of Temperature Control in *Lp* Risk Management**

524 The implementation of a water safety plan is the recommended approach for preventive
525 risk-management related to drinking water (WHO 2011) and temperature control is
526 widely recognized as the first risk mitigation measure for *Legionella* control in hot water
527 distribution systems (HWDS) (Table S1).

528

529 Maintaining sufficient temperatures at all critical points, including the subordinate
530 return loops, and minimizing volumes of uncontrolled temperature in the terminal ends
531 appear essential to a successful system wide thermal control of culturable and VBNC
532 *Legionella*. Most studies report on the results of temperature control based on
533 prevalence measured by culture-based detection methods. Although lower prevalence
534 is generally observed after temperatures are increased, limited efficacies are often
535 reported. An early study observed 50% reduction of tap positivity following an increase
536 in temperature from 45 to 60°C at the water heater outlet, although an elevated
537 number of taps located in patient rooms remained positive (Ezzeddine et al. 1989).
538 Water temperature at the tap ranged between 30 and 56°C after a few minutes of
539 flushing, demonstrating the system's inability to provide elevated temperatures in all
540 areas. A similar reduction in % positive taps from 60-90% to 30-40% was reported in a
541 hospital when water heater temperature was raised from 50 to 65°C, in that case

542 providing temperatures $>50^{\circ}\text{C}$ at most outlets (Blanc et al. 2005). Importantly, the
543 remaining positive outlets were situated in an area with inadequate recirculation. A
544 third field study documented a successful reduction of *Legionella* positive taps from
545 100% to a mean value of 12% maintained over 10 years following the hot water
546 temperature increase from 45 to 65°C (Darelid et al. 2002). This temperature regimen
547 was implemented following an outbreak and resulted in water temperatures between
548 56 and 61°C at the tap after 5 minutes flushing. Recent field studies support the
549 importance of maintaining elevated temperatures at distal locations (estimated by the
550 temperature after 1 minute of flushing), with 4–11% of positive at $T \geq 55^{\circ}\text{C}$ vs 14–82% for
551 $T < 55^{\circ}\text{C}$ (Arvand et al. 2011, Bargellini et al. 2011, Hrubá 2009). Those observations show
552 that the efficiency of thermal inactivation in complex recirculated full scale HWDS is
553 enhanced when temperature exposure is sufficient in all areas of the HWDS. However,
554 significant distal amplification of *Legionella* can occur as evidenced by long term full
555 scale sampling results (Cristina et al. 2014, Serrano-Suarez et al. 2013) and a number of
556 taps may remain positive for *Legionella*.

557

558 The limitations of thermal control in HWDS raise questions on the validity of the existing
559 threshold temperatures of $50\text{--}60^{\circ}\text{C}$. Pioneer work evidenced the consistent
560 susceptibility of 40 *Lp* isolates to temperature, with 1 log reduction achieved in 2.3–5
561 min at 60°C and 8 log reduction after 25 min as estimated by culturability (Stout et al.
562 1986). Recent findings show that elevated temperatures between 55 and 70°C will
563 produce VBNC cells that cannot be detected by culture methods. Laboratory studies

564 conducted on HWDS samples confirm the suppression of culturability at $T \geq 55^\circ\text{C}$ as
565 evidenced by the presence of *Lp* when measured by qPCR and viable qPCR (Lee et al.
566 2011, Mansi et al. 2014). Despite a rapid loss of culturability at temperatures $>55^\circ\text{C}$,
567 some *Lp* strains can resist in the VBNC state for periods of 30-60 minutes at
568 temperatures between 55 and 70°C (Allegra et al. 2008, Allegra et al. 2011, Epalle et al.
569 2014). Furthermore, the development of heat resistant *Lp* strains was observed over
570 time for groups of strains isolated in hospital water systems submitted to periodic
571 extreme temperature (24h @ 65°C a few times a year), while no such resistance was
572 observed for strains isolated from the system where heat shock treatments (70°C 30
573 minutes) were sparingly applied. Finally, the efficacy of thermal disinfection on biofilm,
574 the main reservoir of *Lp* in HWDS (Buse et al. 2014), is at best scarce and reports limited
575 and non-lasting efficacy of 70°C for 2 hours on culturable *Legionella* spp (Saby et al.
576 2005). These findings stress that high temperature regimen provide *Lp* control not *Lp*
577 eradication and the importance of maintaining a constant temperature regimen
578 throughout the system to provide adequate contact time and avoid growth.

579

580 We propose a system wide risk classification to assess risk in a HWDS based on
581 published reports and our findings (Table 2). In addition to monitoring temperature at
582 critical control points, the evaluation criteria also include the percentage of time that
583 temperature is maintained at the hot water production unit or return loops. Indeed,
584 exposure to temperature should be considered instead of temperature alone, as
585 regulated for chemical disinfection (Concentration X Time concept). Subordinate return

586 loop temperatures are used to evaluate the system's heat loss within each sector of the
587 building. Temperature exposure in the subordinate flow and return loop is estimated
588 based on temperatures measured after 1 minute of flushing and serves to determine
589 risk in specific areas. When evaluating the five systems against the proposed risk
590 classification (Table 2), results from the characterization of the HWDS combined with
591 the temperature profiles at point-of-use were good predictors of areas at risk for *Lp*
592 detection (Table 1). In light of these findings and considering the presence of VBNC
593 *Legionella* at temperature ranging between 55-70°C (Epalle et al. 2014), the set points
594 proposed in existing regulations and guidelines and selected for the proposed risk
595 classification approach appear minimal and should be met at all times. The development
596 of heat resistant strains following periodic heat shock also supports the maintenance of
597 a steady thermal preventative inactivation regimen instead of relying on periodic
598 curative thermal shock (Allegra et al. 2011). The apparent limited success of HWDS in
599 large buildings may have been caused by inconsistent maintenance of sufficiently
600 elevated temperatures in all areas of the building because of inadequate recirculation
601 and/or low set-points.

602

603 Regulations and guidelines all recommend the identification of representative sampling
604 points for *Lp* sampling and temperature monitoring at designated control points.
605 However, the rationale for frequency and number of sites for temperature monitoring is
606 not evident and the limited number of proposed control points implies that the HWDS is
607 well balanced. Furthermore, there is little guidance for follow-up action to identify the

608 cause of temperature losses. To remediate this shortfall, a diagnostic flowchart for the
609 initial assessment of *Legionella* risk within an existing HWDS is proposed using
610 temperature measurements and profiles at the water heating unit, return loops and
611 critical points (Fig. 6). We propose a step approach starting from the principal return
612 and flow loop system that indicates the overall system risk level, then moving
613 progressively to the subordinate flow and return loops to identify large building areas or
614 sectors at risk, and finally to the tertiary terminal ends, to identify local issues with
615 defective faucets or showers. The diagnostic flowchart also proposes a staged response
616 in terms of corrective and preventative actions, including *Lp* monitoring. Critical control
617 points, defined as the water heater outlet, the principal return loop and representative
618 at risk points-of-use (not reaching control temperature, farthest from the water heater
619 or serving vulnerable patients) are prioritized for sectors or systems identified at risk by
620 the initial risk assessment (Fig. 6). This step approach can help direct efforts towards
621 high risk areas and optimize resource allocation, especially costly *Lp* monitoring.
622 Nevertheless, an ongoing *Lp* monitoring strategy and schedule should be put into place
623 through a water safety plan once initial assessment is completed and corrective
624 measures have been completed.

625

626 Although temperature control is a central element of risk mitigation, other factors
627 affecting the persistence of *Legionella* in HWDS should be considered in the water
628 safety plan such as: the susceptibility of environmental strains to heat inactivation; the
629 relative importance of terminal volumes not subject to recirculation; the sampling

630 protocol used for *Legionella* monitoring (first volume, flushing, etc.); the presence of
631 biofilm & amoeba and the use of chemical disinfection. The limitations of traditional
632 culture-based methods to detect the presence of *Lp* when in presence of environmental
633 stressors should also be considered when determining a sampling strategy. Although
634 culture is the gold standard and helpful to isolate strains, qPCR can be a valuable tool to
635 monitor changes in a system. An increase in qPCR signal compared to a baseline is
636 indicative of cell growth, even if there is no distinction between viable and dead cells.
637 The relation between *Lp* culture and qPCR results is still not clear, but a recent study by
638 Lee et al. showed qPCR results following the trends of culture in a hot water system,
639 with exceptions for temperatures above 50°C and in the presence of additional
640 disinfection (Lee et al. 2011). These exceptions may be attributed to the impact of
641 temperature on culturability.

642 **4. CONCLUSIONS**

- 643 • A step approach combining temperature monitoring of the hot water
644 distribution system (HWDS) main components and temperature profiling at
645 points-of-use can be used to determine the susceptibility of overall hot water
646 distribution system and specific areas of large buildings to *Legionella*
647 proliferation. When multiple subordinate return loops are present, temperature
648 should be monitored at each subordinate return loop prior to the principal
649 return loop. Monitoring temperature representative points or even all points on

650 a rotating basis (e.g. 20%/year) is time consuming and yet insufficient for rapidly
651 detecting faulty equipment such as defective valves.

- 652 • The impact of faulty thermostatic devices extends far beyond the terminal
653 connecting piping and can affect large areas of buildings, placing significant
654 volumes of hot water at risk. Faulty return valves should be rapidly identified and
655 repaired or replaced. A change observed in results from continuous temperature
656 monitoring of the subordinate return loop can provide useful information to
657 identify the occurrence of a faulty device.
- 658 • Temperature monitoring will help understand the hydraulics, quantify the
659 thermal losses of the recirculating system and identify the distribution columns
660 that need balancing. A systematic diagnostic is necessary to identify areas most
661 at risk in hydraulically unbalanced HWDS or in older buildings where original
662 plans and drawings may not be available or renovations and rearrangements
663 have occurred.
- 664 • Systems assessment and monitoring should also take into account area specific
665 hydraulic conditions within the building, including closed units, low usage and
666 configuration of the overall system.
- 667 • Temperature profiling should be performed at a large number of points
668 confirming the extent (volume) and nature (systemic or distal) of undesirable
669 temperatures in HWDS, guiding *Lp* monitoring decisions. The staged approach
670 based on inexpensive and easily implemented temperature profiling can

671 optimize resources and funds allocation by directing efforts towards high risk
672 areas.

673 • Although necessary, *Lp* monitoring is costly and time-consuming, and should be
674 targeted to enable decision making for infection control. Our staged approach
675 can guide corrective system interventions and serve as a basis to justify
676 preventive risk reduction actions and select sampling points for *Lp* monitoring.

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683 **6. REFERENCES**

684 Allegra, S., Berger, F., Berthelot, P., Grattard, F., Pozzetto, B. and Riffard, S. (2008) Use of
685 flow cytometry to monitor *Legionella* viability. *Applied and Environmental Microbiology*
686 74(24), 7813-7816.

687 Allegra, S., Grattard, F., Girardot, F., Riffard, S., Pozzetto, B. and Berthelot, P. (2011)
688 Longitudinal evaluation of the efficacy of heat treatment procedures against *legionella*
689 *spp.* In hospital water systems by using a flow cytometric assay. *Applied and*
690 *Environmental Microbiology* 77(4), 1268-1275

691 Allen, J.G., Myatt, T.A., Macintosh, D.L., Ludwig, J.F., Minegishi, T., Stewart, J.H.,
692 Connors, B.F., Grant, M.P. and McCarthy, J.F. (2012) Assessing risk of health care-
693 acquired Legionnaires' disease from environmental sampling: the limits of using a strict
694 percent positivity approach. American Journal of Infection Control 40(10), 917-921.

695 Allen, J.G., Gessesse, B., Myatt, T.A., MacIntosh, D.L., Ludwig, J.F., Minegishi, T., Stewart,
696 J.H., Connors, B.F., Grant, M.P., Fragala, M.A. and McCarthy, J.F. (2014) Response to
697 commentary on "Assessing risk of health care-acquired Legionnaires' disease from
698 environmental sampling: The limits of using a strict percent positivity approach".
699 American Journal of Infection Control 42(11), 1250-1253.

700 ASPE (2008) Plumbing Engineering Design Handbook - A Plumbing Engineer's Guide to
701 System Design and Specifications, Volume 4 - Plumbing Components and Equipment.
702 American Society of Plumbing Engineers (ASPE), Chicago, IL.

703 Arvand, M., Jungkind, K. and Hack, A. (2011) Contamination of the cold water
704 distribution system of health care facilities by *Legionella pneumophila*: do we know the
705 true dimension? Eurosurveillance 16(16), 6.

706 AFNOR (2006) NF T90-431/A1 April 2006 Water quality - Detection and enumeration of
707 Legionella spp and Legionella pneumophilia - Method by direct inoculation and after
708 concentration by membrane filtration or centrifugation.

709 Bargellini, A., Marchesi, I., Righi, E., Ferrari, A., Cencetti, S., Borella, P. and Rovesti, S.
710 (2011) Parameters predictive of *Legionella* contamination in hot water systems:

711 Association with trace elements and heterotrophic plate counts. *Water Research* 45(6),
712 2315-2321.

713 Bartram, J., Chartier, Y., Lee, J.V., Pond, K. and Surman-Lee, S. (2007) *Legionella* and the
714 prevention of legionellosis, World Health Organization 2007, Geneva.

715 Best, M., Yu, V.L., Stout, J., Goetz, A., Muder, R.R. and Taylor, F. (1983) *Legionellaceae* in
716 the hospital water-supply: Epidemiological link with disease and evaluation of a method
717 for control of nosocomial legionnaires' disease and Pittsburgh pneumonia. *Lancet*
718 2(8345), 307-310.

719 Blanc, D.S., Carrara, P., Zanetti, G. and Francioli, P. (2005) Water disinfection with ozone,
720 copper and silver ions, and temperature increase to control *Legionella*: seven years of
721 experience in a university teaching hospital. *The Journal of Hospital Infection* 60(1), 69-
722 72.

723 Blokker, E., Vreeburg, J., Schaap, P. and van Dijk, J. (2010) The self-cleaning velocity in
724 practice, WSDA 2010, Tucson, Arizona, USA.

725 Bonetta, S., Ferretti, E., Balocco, F. and Carraro, E. (2010) Evaluation of *Legionella*
726 *pneumophila* contamination in Italian hotel water systems by quantitative real-time PCR
727 and culture methods. *Journal of Applied Microbiology* 108(5), 1576-1583.

728 BSR/ASHRAE (2013) Standard 188P Prevention of legionellosis associated with building
729 water systems. [http://www.r2j.com/wp-](http://www.r2j.com/wp-content/uploads/2013/08/Std188P_3rdPPRDraftFINAL.pdf)
730 [content/uploads/2013/08/Std188P_3rdPPRDraftFINAL.pdf](http://www.r2j.com/wp-content/uploads/2013/08/Std188P_3rdPPRDraftFINAL.pdf)

731 Bujak, J. (2010) Heat consumption for preparing domestic hot water in hospitals. *Energy*
732 and *Buildings* 42(7), 1047-1055.

733 Buse, H.Y., Schoen, M.E. and Ashbolt, N.J. (2012) *Legionellae* in engineered systems and
734 use of quantitative microbial risk assessment to predict exposure. *Water Research* 46(4),
735 921-933.

736 Buse, H.Y., Lu, J., Struewing, I.T. and Ashbolt, N.J. (2014) Preferential colonization and
737 release of *Legionella pneumophila* from mature drinking water biofilms grown on
738 copper versus unplasticized polyvinylchloride coupons *International Journal of Hygiene*
739 and *Environmental Health* 217(2-3), 219–225.

740 CDC (2003) Guidelines for environmental infection control in health-care facilities,
741 Centers for Disease Control and Prevention (CDC), United States Department of Health
742 and Healthcare Infection Control Practices Advisory Committee (HICPAC), Atlanta,
743 Georgia, USA.

744 CSTB (2012) Technical Guide – Controlling the risk of Legionella in sanitary hot water
745 distribution systems - Maîtrise du risque de développement des légionelles dans les
746 réseaux d'eau chaude sanitaire – Faults and recommandations. (In French)

747 CMMQ/ RBQ (2013) Good plumbing practices – Sanitary hot water recirculation system
748 design (part I),
749 [https://www.rbq.gouv.qc.ca/fileadmin/medias/pdf/Publications/francais/conception-](https://www.rbq.gouv.qc.ca/fileadmin/medias/pdf/Publications/francais/conception-boucle-recirculation-eau-chaude.pdf)
750 [boucle-recirculation-eau-chaude.pdf](https://www.rbq.gouv.qc.ca/fileadmin/medias/pdf/Publications/francais/conception-boucle-recirculation-eau-chaude.pdf) (In French).

751 Cristina, M.L., Spagnolo, A.M., Casini, B., Baggiani, A., Del Giudice, P., Brusaferrò, S.,
752 Poscia, A., Moscato, U., Perdelli, F. and Orlando, P. (2014) The impact of aerators on
753 water contamination by emerging gram-negative opportunists in at-risk hospital
754 departments. *Infection Control and Hospital Epidemiology* 35(2), 122-129.

755 Darelid, J., Lofgren, S. and Malmvall, B.E. (2002) Control of nosocomial Legionnaires'
756 disease by keeping the circulating hot water temperature above 55°C: experience from a
757 10-year surveillance programme in a district general hospital. *The Journal of Hospital*
758 *Infection* 50(3), 213-219.

759 Department of Health (DH) and Estates and Facilities Division (2006) Water systems :
760 health technical memorandum 04-01 : The control of *Legionella*, hygiene, "safe" hot
761 water, cold water and drinking water systems. Part B: Operational management,
762 Department of Health (DH), London.

763 DVGW German Technical and Scientific Association for Gas and Water (2004) Technical
764 Rule: Code of Practice W 551. Drinking water heating and drinking water piping systems;
765 technical measures to reduce *Legionella* growth; design, construction, operation and
766 rehabilitation of drinking water installations.

767 Epalle, T., Girardot, F., Allegra, S., Maurice-Blanc, C., Garraud, O. and Riffard, S. (2014)
768 Viable but not culturable forms of *Legionella pneumophila* generated after heat shock
769 treatment are infectious for macrophage-like and alveolar epithelial cells after

770 resuscitation on *Acanthamoeba polyphaga*. *Microbial Ecology*, Published ahead of print
771 July 2014.

772 Ezzeddine, H., Van Ossel, C., Delmee, M. and Wauters, G. (1989) *Legionella spp.* in a
773 hospital hot water system: effect of control measures. *The Journal of Hospital Infection*
774 13(2), 121-131.

775 Health and Safety Executive (HSE) (2013) Legionnaires'disease: Technical guidance. Part
776 2: The control of *Legionella* bacteria in hot and cold water systems, HSE Books, United
777 Kingdom.

778 Hrubá, L. (2009) The colonization of hot water systems by *Legionella*. *Annals of*
779 *Agricultural and Environmental Medicine* 16(1), 115-119.

780 Krojgaard, L., Krogfelt, K., Albrechtsen, H.-J. and Uldum, S. (2011) Detection of *Legionella*
781 by quantitative-polymerase chain reaction (qPCR) for monitoring and risk assessment.
782 *BMC Microbiology* 11(1), 7.

783 Lautenschlager, K., Boon, N., Wang, Y., Egli, T. and Hammes, F. (2010) Overnight
784 stagnation of drinking water in household taps induces microbial growth and changes in
785 community composition. *Water Research* 44(17), 4868-4877.

786 Lee, J.V., Lai, S., Exner, M., Lenz, J., Gaia, V., Casati, S., Hartemann, P., Lück, C., Pangon,
787 B., Ricci, M.L., Scaturro, M., Fontana, S., Sabria, M., Sánchez, I., Assaf, S. and Surman-
788 Lee, S. (2011) An international trial of quantitative PCR for monitoring *Legionella* in
789 artificial water systems. *Journal of Applied Microbiology* 110(4), 1032-1044.

790 Lin, Y.E., Stout, J.E. and Yu, V.L. (2011) Controlling *Legionella* in hospital drinking water:
791 an evidence-based review of disinfection methods. Infection Control and Hospital
792 Epidemiology 32(2), 166-173.

793 Lipphaus, P., Hammes, F., Kotsch, S., Green, J., Gillespie, S. and Nocker, A. (2014)
794 Microbiological tap water profile of a medium-sized building and effect of water
795 stagnation. Environmental Technology 35(5-8), 620-628.

796 Mansi, A., Amori, I., Marchesi, I., Proietto, A.R., Marcelloni, A.M., Ferranti, G., Magini, V.,
797 Valeriani, F. and Borella, P. (2014) *Legionella* spp. survival after different disinfection
798 procedures: Comparison between conventional culture, qPCR and EMA–qPCR.
799 Microchemical Journal 112, 65-69.

800 Martinelli, F., Caruso, A., Moschini, L., Turano, A., Scarcella, C. and Speziani, F. (2000) A
801 Comparison of *Legionella pneumophila* occurrence in hot water tanks and Instantaneous
802 devices in domestic, nosocomial, and community environments. Current Microbiology
803 41(5), 374-376.

804 Moritz, A.R. and Henriques, F.C. (1947) Studies of thermal injury. II. The relative
805 importance of time and surface temperature in the causation of cutaneous burns. The
806 American Journal of Pathology 23(5), 695-720.

807 Napoli, C., Iatta, R., Fasano, F., Marsico, T. and Montagna, M.T. (2009) Variable bacterial
808 load of *Legionella* spp. in a hospital water system. Science of the Total Environment
809 408(2), 242-244.

810 Pierre, D., Stout, J.E. and Yu, V.L. (2014) Editorial commentary: Risk assessment and
811 prediction for health care-associated Legionnaires' disease: Percent distal site positivity
812 as a cut-point. American Journal of Infection Control 42(11), 1248-1250.

813 République Française (2005) Ministerial Order, November 30th 2005, modifying the
814 Ministerial order of June 23rd 1978 on sanitary hot water production and distribution
815 stationary installations for residential, work or public buildings,
816 http://www.sante.gouv.fr/fichiers/bo/2011/11-01/ste_20110001_0100_0130.pdf (In
817 French).

818 République Française (2010a) Ministerial Order February 1st 2010, Legionella
819 surveillance in sanitary hot water production, storage and distribution installations,
820 <http://www.legifrance.gouv.fr/affichTexte.do?cidTexte=JORFTEXT000021795143&dateT>
821 [exte=&categorieLien=id](http://www.legifrance.gouv.fr/affichTexte.do?cidTexte=JORFTEXT000021795143&dateT). (In French)

822 République Française (2010b) Circular DGS/EA4/2010/448, December 21st 2010
823 regarding the implementation of February 1st 2010 Ministerial Order Legionella
824 surveillance in sanitary hot water production, storage and distribution installations,
825 http://www.sante.gouv.fr/fichiers/bo/2011/11-01/ste_20110001_0100_0130.pdf. (In
826 French)

827 Saby, S., Vidal, A. and Sutty, H. (2005) Resistance of *Legionella* to disinfection in hot
828 water distribution systems. Water Science and Technology 52(8), 15-28.

829 Serrano-Suarez, A., Dellunde, J., Salvado, H., Cervero-Arago, S., Mendez, J., Canals, O.,
830 Blanco, S., Arcas, A. and Araujo, R. (2013) Microbial and physicochemical parameters
831 associated with *Legionella* contamination in hot water recirculation systems.
832 Environmental Science and Pollution Research International 20(8), 5534-5544.

833 Stout, J.E., Best, M.G. and Yu, V.L. (1986) Susceptibility of members of the family
834 *Legionellaceae* to thermal stress: implications for heat eradication methods in water
835 distribution systems. Applied Environmental Microbiology 52(2), 396-399.

836 Stout, J.E., Muder, R.R., Mietzner, S., Wagener, M.M., Perri, M.B., DeRoos, K., Goodrich,
837 D., Arnold, W., Williamson, T., Ruark, O., Treadway, C., Eckstein, E.C., Marshall, D.,
838 Rafferty, M.E., Sarro, K., Page, J., Jenkins, R., Oda, G., Shimoda, K.J., Zervos, M.J., Bittner,
839 M., Camhi, S.L., Panwalker, A.P., Donskey, C.J., Nguyen, M.H., Holodniy, M., Yu, V.L. and
840 the Legionella Study Group (2007) Role of environmental surveillance in determining the
841 risk of hospital-acquired legionellosis: a national surveillance study with clinical
842 correlations. Infection Control and Hospital Epidemiology 28(7), 818-824.

843 WHO (2011) Water Safety in Buildings. Resource for the Development of Training and
844 Information Material. WorldHealth Organization, Geneva.

845 Yee, R.B. and Wadowsky, R.M. (1982) Multiplication of *Legionella pneumophila* in
846 unsterilized tap water. Applied and Environmental Microbiology 43(6), 1330-1334

847 Yu, Z. and Mohn, W.W. (1999) Killing two birds with one stone: simultaneous extraction
848 of DNA and RNA from activated sludge biomass. Canadian Journal of Microbiology 45(3),
849 269-272.

850

851 **Figure legends:**

852 Fig. 1: Hot water distribution system general schematic including temperature control
853 points. Three different types of vertical and horizontal distribution systems are
854 represented: ① recirculation before the last tap; ② recirculation connected after each
855 device; ③ recirculation connected after the last device.

856 Fig. 2: Examples of continuous temperature monitoring at water heater outlet for each
857 studied system (black line) and at the return loop for system 4 and 5 (blue line). No
858 continuous monitoring data was available for systems 1 to 3. Date format is MM/DD/YY.

859 Fig. 3: Heat loss during stagnation of hot water in 1.25 cm diameter copper pipes with
860 and without insulation at room temperature.

861 Fig. 4: Hot water temperature profiles at points-of-use as a function of volume for a)
862 systems 1, 2 and 3 grouped, n = 7; b) system 4, n = 7; c) system 5, n = 36. Mean
863 temperature at the hot water production unit outlet and at the return loop are shown
864 for each system.

865 Fig. 5: Variability of *L. pneumophila* concentration measured by qPCR (a) in system 4, for
866 repeat sampling events without prior stagnation (n = 5, Jan-Oct 2013) (b) in system 5,
867 after different water stagnation times for tap A (light gray) and tap B (dark gray) (n = 2,
868 Nov-Oct 2012).

869

870 Fig. 6: Diagnostic flowchart for the initial assessment of *Legionella* risk in an existing
871 HWDS.

872 Table 1: HWDS Systems characterization through control points temperature and
873 microbiological measurements

874 Table 2: Proposed risk classification based on temperature control points

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