

Aluminum effects on marine phytoplankton: implications for a revised Iron

Hypothesis (Iron-Aluminum Hypothesis)

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1 **Abstract**

2 In contrast to substantial studies and established knowledge of aluminum (Al) effects
3 (mainly toxicity) on freshwater organisms and terrestrial plants, and even on human
4 health, only a few studies of Al effects on marine organisms have been reported, and our
5 understanding of the role of Al in marine biogeochemistry is limited. In this paper, we
6 review the results of both field and laboratory experiments on the effects of Al on marine
7 organisms, including Al toxicity to marine phytoplankton and the beneficial effects of Al
8 on marine phytoplankton growth, and we discuss possible links of Al to the biological
9 pump and the global carbon cycle. We propose a revised Iron (Fe) Hypothesis, i.e., the
10 Fe-Al Hypothesis that introduces the idea that Al as well as Fe play an important role in
11 the glacial-interglacial change in atmospheric CO₂ concentrations. We propose that Al
12 could not only facilitate Fe utilization, dissolved organic phosphorus utilization and
13 nitrogen fixation by marine phytoplankton, enhancing phytoplankton biomass and carbon
14 fixation in the upper oceans, but also reduce the decomposition and decay of biogenic
15 matter. As a result, Al allows potentially more carbon to be exported and sequestered in
16 the ocean depths through the biological pump. We also propose that Al binds to
17 superoxide to form an Al-superoxide complex, which could catalyze the reduction of
18 Fe(III) to Fe(II) and thus facilitate Fe utilization by marine phytoplankton and other
19 microbes. Further ocean fertilization experiments with Fe and Al are suggested, to clarify

1 the role of Al in the stimulation of phytoplankton growth and carbon sequestration in the
2 ocean depths.

3

4 **Keywords:** aluminum; marine phytoplankton; Iron Hypothesis; Fe-Al Hypothesis;
5 beneficial effects; biological pump

6

1 **1 Introduction**

2 Aluminum (Al) is the most abundant metallic element in the Earth's crust (Taylor
3 1964), it is widespread in the environment, and is present in every cell of every organism
4 (Exley and Mold 2015). Paradoxically, no established biological function of Al has yet
5 been found.

6 Aluminum can be toxic to aquatic organisms and terrestrial plants, and it has even
7 been related to some human diseases (Gensemer and Playle 1999; Macdonald and Martin
8 1988; Yokel 2000). In contrast to substantial studies and established knowledge of Al
9 effects on organisms in acid and neutral pH freshwater, only a few studies of Al effects
10 on marine organisms in seawater have been reported. Our understanding of the role of Al
11 in marine biogeochemistry is strictly limited.

12 Many studies focus on the scavenging effects of marine organisms on the distribution
13 of Al in seawater, as marine organisms, especially microbes, can adsorb Al on their
14 surface or absorb it into cells, thus decreasing dissolved Al concentrations in seawater
15 (e.g., Liu et al. 2017b; Moran and Moore 1988; Ren et al. 2011). In contrast, the effects
16 of Al on marine organisms are usually unnoticed. More than five decades ago, Menzel et
17 al. (1963) reported that Al enrichment could increase primary productivity in some
18 waters in the Sargasso Sea. Subsequently, over the past half century, sporadic studies
19 have reported the influences of Al on marine organisms including the early life stages of
20 oysters, mussels, crabs, urchins and cnidarians, polychaetes, and marine algae

1 (Supplement material Table S1). All these studies provide a preliminary overall notion
2 that the Al effects on marine organisms might be very different, and more diverse,
3 compared to its mainly toxic interactions with organisms in freshwater. These results also
4 inspired us to think that Al may play an important but unrecognized role in the global
5 carbon cycle through influencing marine organisms, especially marine phytoplankton.

6
7 To understand the role of Al in marine biogeochemistry, we review 1) the
8 bioavailability of Al to marine organisms, 2) Al toxicity to marine organisms, 3)
9 beneficial effects of Al on marine phytoplankton growth, 4) the possible links of Al to the
10 biological pump and the global carbon cycle, and 5) the relationship of Al with climate
11 change and the Iron Hypothesis. We propose a revised Fe-Al Hypothesis to recognize the
12 potentially important role of Al in the global carbon cycle and climate change over
13 geological time scales.

14

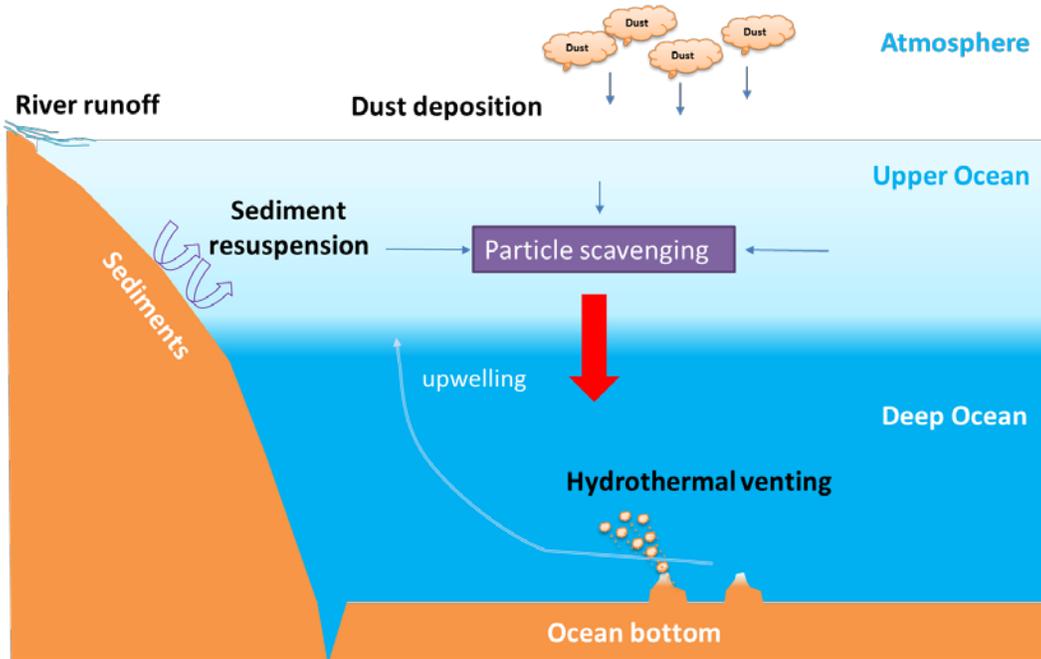
15 **2 Bioavailability of Al to marine organisms**

16 **2.1 Aluminum sources in natural seawater**

17 River runoff, atmospheric dust deposition, and sediment resuspension are usually
18 considered the main sources of Al in natural seawater (Maring and Duce 1987; Orians
19 and Bruland 1986; van Hulst et al. 2013) (Fig. 1). Aluminum is transported with river
20 runoff into estuaries and their adjacent coastal waters, and with the deposition of
21 atmospheric dust into offshore seawater and the open ocean. Sediment resuspension

1 could also release Al trapped in the bottom (biogenic and abiotic) sediments into the
2 overlying seawater. Recent studies show that hydrothermal venting is another potentially
3 important source of Al in the ocean (Fig. 1). Hydrothermal vents in mid-ocean ridges,
4 such as the southern East Pacific Rise, are thought to contribute an important flux of Al
5 into the adjacent deep ocean (Elderfield and Schultz 1996), and the Al could be
6 transported several thousand kilometers westward across the Southern Ocean (Measures
7 et al. 2015; Resing et al. 2015). In addition, anthropogenic activities, such as coastal
8 mining and dredging operations, discharges of liquid wastes associated with Al
9 production, and the use of Al in sacrificial anodes, could significantly increase local
10 inputs of Al into natural seawater (Gillmore et al. 2016).

11



12

13 **Figure 1.** Schematic diagram of natural sources of Al in the ocean.

1

2 **2.2 Solubility of Al in seawater**

3 The solubility of Al is pH-dependent. Like most trivalent metals, Al is more soluble
4 in acidic and basic solutions with a minimum in circumneutral solutions (Millero 2009).
5 Savenko and Savenko (2011) examined the solubility of Al in artificial seawater with a
6 salinity of 35 at 25 °C and over a pH range from 7.4 to 8.2. They found that the solubility
7 ranged from 1.9 to 6.2 μM , with a minimum from pH 8.05 to 8.10. An Al solubility of
8 about 10.5 μM in artificial seawater has also been reported (Machill et al. 2013).

9 The solubility of Al in natural seawater is significantly higher than that in artificial
10 seawater. Willey (1975) reported that the Al solubility in filtered offshore seawater
11 sampled in the Sargasso Sea was 18.5 μM (500 $\mu\text{g/L}$) at 2 °C and demonstrated that Al
12 could be removed from seawater by coprecipitation with added silicic acid. He concluded
13 that levels of dissolved Al found in seawater would vary as a function of the
14 concentration of dissolved silica. Another two studies reported Al solubility values for
15 filtered coastal seawater of 18.5 μM (500 $\mu\text{g/L}$) at 22 °C, and 24 μM (650 $\mu\text{g/L}$) at 28 °C,
16 respectively (Angel et al. 2016; Trenfield et al. 2015). Owing to slow rates of
17 precipitation and transformation of Al solid phases, dissolved Al concentrations up to
18 1250 $\mu\text{g/L}$ (46.3 μM) could also persist for several days (Angel et al. 2016). The much
19 higher solubility of Al in natural seawater than in artificial seawater might be partly due
20 to the complexation of Al by strong organic ligands in natural seawater, as there is

1 evidence indicating that siderophores, the high-affinity iron-chelating molecules secreted
2 by microorganisms such as bacteria, could complex Al (Santos 2008).

3

4 **2.3 Speciation of dissolved Al in seawater**

5 At present, there are no analytical methods to measure the ionic and neutral dissolved
6 forms of Al in seawater (Gillmore et al. 2016), and our knowledge about Al speciation in
7 seawater is based almost entirely on thermodynamic calculations. Without considering
8 the influence of organic ligands, calculations with chemical equilibrium models show that
9 aluminate (Al(OH)_4^-), aluminum trihydroxide (Al(OH)_3) and Al(OH)_2^+ are the dominant
10 Al species in alkaline seawater, although the proportions of the three species relative to
11 total dissolved Al in seawater vary among the different models (Table 1). Based on the
12 Miami Interactions Model (Pierrot and Millero 2017), which is designed for calculating
13 trace metal speciation in seawater, aluminate and aluminum trihydroxide are the
14 dominant species in the pH range (7.4 to 9.0) of natural seawater (data not shown); this
15 result differs from the speciation proposed by Savenko and Savenko (2011), where
16 Al(OH)_2^+ is the primary form of Al occurrence in seawater at $\text{pH} < 8.05$ and Al(OH)_3
17 prevails within the narrow range from pH 8.05 to 8.10. Aluminum can also react with
18 silicic acid but this reaction tends to produce colloidal and particulate
19 hydroxyaluminosilicates rather than dissolved species (Beardmore et al. 2016; Exley et al.
20 2012).

1 Despite the lack of agreement concerning the exact proportions of the various
 2 $\text{Al(OH)}_x^{(3-x)+}$ complexes in seawater, the species of dissolved Al in seawater are clearly
 3 very different from the main Al species in neutral and weakly acid freshwater. Generally,
 4 Al^{3+} , Al(OH)^{2+} , Al(OH)_2^+ , and Al-fulvic are the dominant dissolved Al species in natural
 5 freshwater (Campbell et al. 1992; Driscoll and Schecher 1990).

6

7 **Table 1** Speciation of dissolved Al in seawater calculated by different models at pH =
 8 8.1 and 25 °C. Seawater composition used for the models was documented by
 9 Pierrot & Millero (2017). The total Al concentration was set as 100 nM.

Species	Miami Interactions Model	MINEQL 5.0 ^a Model	Visual MINTEQ Model
Al(OH)_2^+	0.36%	< 0.03%	< 0.04%
Al(OH)_3	35.3%	0.5%	1.1%
Al(OH)_4^-	64.4%	99.5%	98.9%

10 ^a The MINEQL5.0 model predicted that ~61% of the total 100 nM Al would
 11 precipitate as diaspore; data in the table show the proportions in the remaining
 12 dissolved Al.

13

14 **2.4 Dissolved Al concentrations in natural seawater**

1 Aluminum concentrations in natural seawater vary significantly, ranging from
2 sub-nanomolar (nM) in the open ocean (Kaupp et al. 2011; Middag et al. 2011;
3 Planquette et al. 2009) to micromolar (μM) in estuaries. The highest Al concentrations in
4 natural seawater occur in the estuaries, where typical Al concentrations range from 10 to
5 several hundred nM. Concentrations as high as μM can be observed in brackish waters
6 with low salinity in the riverine end of estuaries (Angel et al. 2016; Hall et al. 1999;
7 Zhang et al. 1999), although, strictly speaking, the salinity at the riverine end is too low
8 to be considered as seawater. Dissolved Al concentrations in coastal seawater and in
9 ocean basins have been summarized by several studies (Angel et al. 2016; Han et al. 2008;
10 van Hulst et al. 2013). These studies indicate that typical Al concentrations in coastal
11 waters range from 10 to 100 nM, whereas typical Al concentrations in the open ocean
12 range from sub-nM to tens of nM. The mean Al concentrations in surface seawaters in
13 different ocean basins, such as the North Atlantic, South Atlantic, Pacific, Southern
14 Ocean, Indian Ocean and Mediterranean Sea, were reported to range from 2.2 nM to 65.2
15 nM (Han et al. 2008). Clearly, Al concentrations in most of the ocean are very low, i.e., at
16 trace levels.

17

18 **2.5 Accumulation and uptake of Al by marine organisms**

19 One of the reasons for the relatively low Al concentrations in natural seawater is that
20 dissolved Al in seawater is usually quickly scavenged by particles, such as marine

1 plankton (Fig. 1). Over the last several decades, a number of studies have reported that
2 marine plankton, especially marine phytoplankton such as diatoms, could remove or
3 scavenge dissolved Al in seawater through adsorption on their surface or/and absorption
4 into their cells (Liu et al. 2017b; Moran and Moore 1988; Ren et al. 2011).

5 Consistent with the idea that Al exists in every cell of every organism (Exley and
6 Mold 2015), marine plankton can also accumulate large amounts of Al, which is usually
7 the most abundant trace metal in marine plankton (Collier and Edmond 1984; Ebihara et
8 al. 2006; Kuss and Kremling 1999; Martin and Knauer 1973). In particular, marine
9 diatoms scavenge dissolved Al in seawater efficiently, and there is evidence that marine
10 diatoms can incorporate Al into their siliceous frustules (Gehlen et al. 2002; Koning et al.
11 2007; Machill et al. 2013; Moran and Moore 1988; Stoffyn 1979). Aluminum has been
12 shown to bind to the cell walls (frustules) of marine diatoms (e.g., *Porosira glacialis*,
13 *Thalassiosira nordenskiöldii*, *Lauderia annulata*, and *Stephanopyxis turris*), and Al/Si
14 ratios in diatom frustules ranging from 1:10,000 to 1:15 have been reported (Beck et al.
15 2002; Machill et al. 2013; Van Bennekom et al. 1991). To our knowledge, there is only
16 one published study reporting that Al could be transported into the cells of three marine
17 diatoms (*Nitzschia closterium*, *Skeletonema coasatum* and *Thalassiosira weissflogii*).
18 However, in this laboratory study (Wang et al. 2013), truly intracellular Al was not
19 distinguished from the fraction of Al that was strongly bound to the cell surface (i.e., the
20 siliceous cell wall) and could not be removed by washing solutions designed to remove

1 metals weakly adsorbed onto the cell surface. Our own unpublished data indicate that the
2 Al internalization flux for a model marine diatom (*Thalassiosira weissflogii*), normalized
3 to the exposure concentration, was 0.033 ± 0.013 nmol/m²/min/nM, and the assimilated
4 Al was distributed in descending order of importance among the following subcellular
5 fractions: granules ($68.7 \pm 5.4\%$), debris ($17.3 \pm 3.6\%$), organelles ($11.9 \pm 2.1\%$),
6 heat-stable peptides ($1.8 \pm 0.2\%$), and heat-denaturable proteins ($0.4 \pm 0.0\%$).

7 Exley and Mold (2015) identified five major routes by which Al could traverse cell
8 membranes or epi-/endo-thelia, including paracellular transport, transcellular transport,
9 active transport, ion channels and adsorptive or receptor-mediated endocytosis. They
10 suggested that Al could be assimilated in this manner in a number of forms: 1) free
11 solvated trivalent cation, 2) low molecular weight, neutral, soluble complexes, 3) high
12 molecular weight, neutral, soluble complexes, 4) low molecular weight, charged, soluble
13 complexes, and 5) nano and micro-particulates. However, for trivalent metals such as Al,
14 these uptake mechanisms are still speculative, as a result of insufficient study and
15 technical challenges. The aqueous coordination chemistry of Al is complex (e.g., slow
16 kinetics and metastable poly-Al forms), it binds strongly to biological surfaces, no
17 affordable and suitable radioisotopes are available and sensitive analytical techniques
18 seldom exist (Crémazy et al. 2013; Rengel 1996; Taylor et al. 2000). To our knowledge,
19 there are no studies on the uptake kinetics of Al assimilation by marine organisms, let

1 alone on the mechanisms of Al transport into marine phytoplankton cells. Overall, we
2 know very little about the bioavailability of Al to marine organisms.

3

4 **3 Al toxicity to marine organisms**

5 There are very few studies reporting the effects (including toxicity) of Al on marine
6 organisms. To the best of our knowledge, only 29 published studies (excluding studies of
7 the influence of Al on diatom frustules) report on the effects of Al on marine organisms
8 (supplemental material Table S1); more than half of the studies (16) have been published
9 since 2010, and only 16 of the studies were intentionally designed to test the acute
10 toxicity of Al to marine organisms or organisms in brackish waters.

11 Tolerance of high concentrations of Al by marine animals has been highlighted by a
12 number of studies. The toxicity of Al to the early life stages (embryonic development,
13 fertilization rates, etc.) of marine invertebrates such as oysters, mussels, crabs, urchins,
14 cnidarians and polychaetes has been relatively substantially covered in early (Calabrese
15 et al. 1973; Macdonald et al. 1988; Pagano et al. 1996; Petrich and Reish 1979; Wilson
16 and Hyne 1997) and recent studies (Caplat et al. 2010; Golding et al. 2015; Negri et al.
17 2011). The Al toxicity to copepods and fish in brackish water, and to fish in seawater, has
18 also been documented (Bengtsson 1978; Golding et al. 2015; Teien et al. 2006). These
19 studies demonstrate that Al toxicity only occurs at μM concentrations or above for most
20 of the tested animal species (supplemental material Table S1).

1 Aluminum toxicity to marine phytoplankton has only been intentionally tested in
2 recent years (Gillmore et al. 2016; Golding et al. 2015; Harford et al. 2011; Saçan et al.
3 2007; Xie et al. 2015). The influence of Al on the growth, iron (Fe) uptake and
4 physiology of marine phytoplankton has also been reported in other studies (Liu et al.
5 2018; Santana-Casiano et al. 1997; Shi et al. 2015; Stoffyn 1979; Vrieling et al. 1999;
6 Zhou et al. 2016). Among all the 13 marine phytoplankton species tested (Table S1), the
7 growth of only one diatom *Ceratoneis closterium* was reported to be inhibited by
8 environmentally relevant Al concentrations (higher than about 10 µg/L or 0.37 µM) in
9 one study (Golding et al. 2015), but its growth was reported to be stimulated by low Al
10 concentrations (< 60 µg/L or < 2.2 µM) in another study (Gillmore et al. 2016). Similarly,
11 the growth and nitrogen fixation of the cyanobacterium *Crocospaera watsonii* were
12 inhibited by a low Al concentration (200 nM) in a phosphorus-sufficient seawater
13 medium (Liu et al. 2017a), but they were stimulated by the same Al concentration in the
14 same seawater medium but with insufficient phosphorus (Liu et al. 2018). Overall, based
15 on the existing published data, environmentally relevant Al concentrations will not be
16 toxic to most marine phytoplankton species in natural seawater.

17 It should be pointed out here that recent studies show that the Al effects on marine
18 phytoplankton are closely related to the nutrient status of the algal growth medium (Liu et
19 al. 2017a; 2018; Zhou et al. 2016). In future studies, nutrient conditions in the medium
20 used for Al toxicity tests should be considered as a potential factor influencing the

1 toxicity test results. We speculate that different background nutrient concentrations in the
2 seawater used for preparing the test media might account for the inconsistent toxicity
3 results for *Ceratoneis closterium* (Golding et al. 2015; Gillmore et al. 2016).

4 Consistent with the above discussion and based on toxicity tests of 11 species from 6
5 taxonomic groups, Golding et al. (2015) estimated that 2.1 µg/L and 24 µg/L of Al (i.e.,
6 78 and 890 nM) would be safe for 99% and 95% of marine species, respectively.

7 Although our understanding of Al effects on marine organisms is far from sufficient,
8 based on our present knowledge about Al concentrations in natural seawater (discussed in
9 Section 2.4), we would expect that natural levels of Al in most of the ocean will show no
10 toxicity to marine organisms living there.

11

12 **4 Beneficial effects of Al on marine phytoplankton growth**

13 **4.1 Enhanced growth of marine phytoplankton in the presence of Al**

14 In contrast to Al toxicity, there are several studies reporting that Al could be
15 beneficial to the growth of marine phytoplankton, “growth” being defined in the present
16 context as the exponential growth rate and/or biomass yield.

17 Since Menzel et al. (1963) first reported that Al enrichment increased the primary
18 productivity in some waters in the Sargasso Sea, additional evidence indicating the
19 beneficial effects of Al on marine phytoplankton growth has been sporadically published.
20 Stoffyn (1979) reported the stimulatory effects of Al on the growth of a marine diatom

1 *Skeletonema costatum* during a study on diatom control of dissolved Al in seawater.
2 Twenty years later, it was reported that enrichments with Al (0.1 to 10 μ M) increased
3 growth of a pennate diatom *Navicula salinarum* (Vrieling et al. 1999). Enhanced growth
4 of other marine phytoplankton species (*Dunaliella tertiolecta*, *Tetraselmis* sp., and
5 *Ceratoneis closterium*) in the presence of Al was unexpectedly observed during tests of
6 Al toxicity to marine phytoplankton (Gillmore et al. 2016; Golding et al. 2015; Saçan et
7 al. 2007). Similar stimulatory effects of Al enrichment on marine phytoplankton growth
8 (e.g., *Synechococcus* sp., *Thalassiosira weissflogii* and *Crocospaera watsonii*) were
9 reported in laboratory studies (Leleyter et al. 2016; Liu et al. 2018; Shi et al. 2015; Shi et
10 al. 2016; Zhou et al. 2016). Enhanced growth of marine phytoplankton (e.g., Diatoms,
11 *Trichodesmium*, photosynthetic picoeukaryotes, unicellular diazotrophic cyanobacterium
12 group A, *Synechococcus* and *Prochlorococcus*) in the presence of added Al has also been
13 observed in field studies (Liu et al. 2018; Shi et al. 2016; Zhou et al. 2017). Our
14 unpublished data indicate that Al enrichment (40 nM and 200 nM) enhanced the yield of
15 all the tested diatoms, namely *Thalassiosira oceanica*, *T. pseudonana*, *Phaeodactylum*
16 *tricornutum* and *Chaetoceros muelleri*, in phosphorus-deficient Aquil medium (Zhou et
17 al. unpublished manuscript).

18

19 **4.2 Enhanced nitrogen fixation in the presence of Al**

1 Previous studies indicate that Al is linked to nitrogen fixation and nitrogen-fixers.
2 Positive correlations between dissolved Al, nitrogen fixation and *Trichodesmium*
3 abundance have been reported in the Atlantic Ocean (Moore et al. 2009; Rijkenberg et al.
4 2011). In these studies Al was treated only as an indicator of terrigenous dust (/Fe) input,
5 and the roles of Al in influencing nitrogen fixation and nitrogen-fixers were usually
6 unnoticed. In fact, *Trichodesmium* could not only accumulate a large amount of Al (e.g.,
7 2.6 $\mu\text{mol}/\mu\text{g}$ chlorophyll-*a*) (Moore et al. 2009; Tovar-Sanchez et al. 2006), but it also
8 contains a gene of unknown function involved in Al resistance (da Cunha et al. 2009).

9 A recent study shows that enhanced nitrogen fixation and increased growth of
10 nitrogen-fixers occurs in the presence of Al. By adding Al to natural seawater collected
11 from the South China Sea, a large subtropical and tropical marginal sea, we observed
12 enhanced growth of *Trichodesmium*, and increased nitrogen fixation by the whole
13 plankton community (Zhou et al. 2017). Another study indicated that addition of a low
14 level of Al (200 nM) to natural seawater in the South China Sea could not only stimulate
15 nitrogen fixation by the whole plankton community, but could also increase the growth
16 and the *nifH* gene expression of two nitrogen-fixers (*Trichodesmium* spp. and unicellular
17 diazotrophic cyanobacterium group A (Liu et al. 2018). Stimulatory effects of Al on the
18 growth and nitrogen fixation of the unicellular diazotrophic cyanobacterium
19 *Crocospaera watsonii* in phosphorus-deficient Aquil medium have also been confirmed
20 in a laboratory study (Liu et al. 2018).

21

1

2 **4.3 Enhanced uptake of Fe in the presence of Al**

3 Another beneficial effect of Al on marine phytoplankton growth is that some marine
4 diatoms increase their Fe uptake in the presence of Al. Santana-Casiano et al. (1997)
5 reported that the Fe uptake by *Thalassiosira weissflogii* increased in the presence of Al.
6 They suggested that an increase of labile Fe could be one of the reasons for increased Fe
7 uptake by the diatom in the presence of Al. However, they recognized that the
8 mechanism for the transformation reactions from stable Fe-organic complexes, inorganic
9 crystalline Fe hydrolysis products and small Fe colloids to labile Fe was not clear.

10 New understanding of Fe uptake by marine microorganisms, and the catalytic role of
11 Al in the reduction of Fe(III) to Fe(II), suggests a new perspective on why the presence of
12 Al could increase Fe uptake by marine phytoplankton. On the one hand, extracellular
13 reduction of Fe (III) to Fe (II) is considered a prerequisite for Fe uptake by most
14 phytoplankton (Shaked et al. 2005), and an Fe-acquisition strategy allowing other
15 microbes (such as denitrifiers or anammox-capable species) to succeed in regions where
16 the concentrations of bioavailable Fe are exceedingly low (Kondo and Moffett 2015;
17 Moffett et al. 2007). Extracellular reduction of Fe(III) complexes by biogenic superoxide
18 is an important way of producing Fe(II) (Rose and Waite 2005). Besides photochemical
19 production of superoxide in the marine environment, many marine organisms including
20 diatoms, cyanobacteria and heterotrophic bacteria can produce superoxide, which is

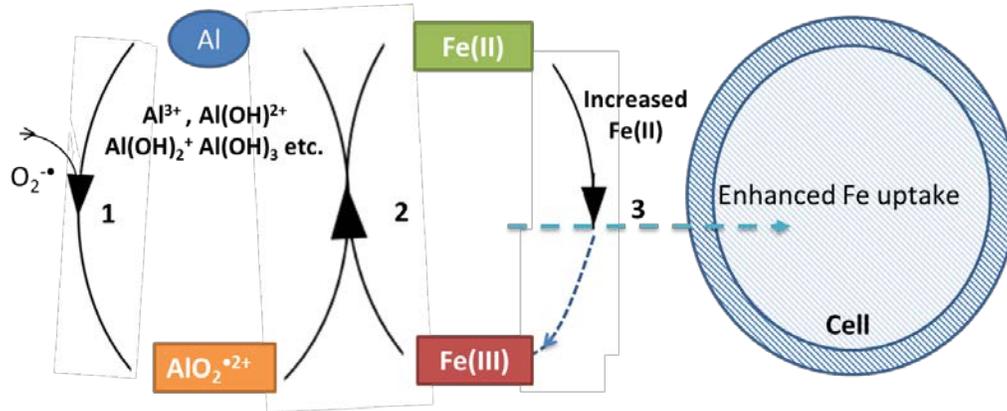
1 capable of reducing a wide range of forms of Fe(III) under typical environmental
2 conditions, such as those found in many marine systems (Diaz et al. 2013; Hansel et al.
3 2016; Rose 2012; Schneider et al. 2016).

4 On the other hand, Al can potentially facilitate the reduction of Fe(III) to Fe(II) and
5 increase Fe availability to marine organisms. First, Al has an intrinsic high potential to
6 stabilize the superoxide anion (Exley 2004; Mujika et al. 2011). This could increase the
7 lifetimes of superoxide long enough to enable it diffuse well away from the site of its
8 production, and influence the reduction of Fe(III) on a spatial scale that is biologically
9 significant (Rose 2012). Second, Al could bind with superoxide to form an Al-superoxide
10 complex, catalyzing the reduction of Fe(III) to Fe(II), and increasing the concentration of
11 Fe(II) (Exley 2004; Khan et al. 2006; Mujika et al. 2011; Ruipérez et al. 2012); the result
12 would be an increase in labile and readily internalized Fe (Fig. 2).

13 Accordingly, we hypothesize that Al can facilitate Fe uptake by marine
14 phytoplankton and other microbes through forming an Al-superoxide complex, catalyzing
15 the reduction of Fe(III) to Fe(II) and increasing Fe bioavailability. The superoxide could
16 be biogenic in origin or from other external sources, as discussed above.

17

18



1

2 **Figure 2** Schematic diagram illustrating how Al could facilitate Fe uptake by marine

3 phytoplankton and other microbes. 1. Al binds with superoxide to form an

4 Al-superoxide complex (e.g. AlO₂^{•2+}); 2. Al-superoxide catalyzes the reduction

5 of Fe (III) to Fe (II); 3. Increased bioavailability of Fe (II) results in enhanced

6 Fe uptake by phytoplankton (or other microbial) cells. The thermodynamic

7 tendency to form Al-superoxide is Al species-dependent, and it decreases in the

8 following order: Al³⁺, Al(OH)²⁺, Al(OH)₂⁺, Al(OH)₃ (Ruipérez et al. 2012).

9

10 **4.4 Aluminum facilitates dissolved organic phosphorus (DOP) utilization by** 11 **marine phytoplankton**

12 Zhou et al. (2016) suggested that the Al-related enhanced utilization of DOP could

13 account for the increased yield of marine phytoplankton that was observed in the

14 presence of Al under conditions of phosphorus limitation. To examine why Al

15 enrichment could stimulate the yield of *Thalassiosira weissflogii* in phosphorus-deficient

16 media, but did not influence diatom growth and yield in phosphorus-sufficient media,

1 they demonstrated that the Al-treated *T. weissflogii* cells utilized DOP at a higher rate
2 than the control cells under conditions of phosphorus limitation. Their results indicate
3 that Al enrichment led to a higher proportion of alkaline phosphatase activity (APA)
4 being associated with the diatom cell surface, and a lower proportion of APA in dissolved
5 form. More interestingly, the Al-treated diatom cells could use cellular APA more
6 efficiently to utilize DOP than could the control cells, as the ratio of DOP uptake rate to
7 cellular APA was significantly higher for the Al-treated cells than for the control cells.
8 The increased yield of other diatom species (*Thalassiosira oceanica*, *Thalassiosira*
9 *pseudonana*, *Phaeodactylum tricornutum*, *Chaetoceros muelleri*) and growth of one
10 nitrogen-fixing cyanobacterium (*Crocospaera watsonii*) in the presence of Al in
11 phosphorus-deficient Aquil media could also be related to the enhanced utilization of
12 DOP (Zhou et al. unpublished manuscript and Liu et al. 2018).

13 It should be pointed out that, despite the demonstration of enhanced utilization of Fe
14 and DOP by marine phytoplankton in the presence of Al, we are still far from a full
15 understanding of the mechanisms underlying the beneficial effects of Al on marine
16 phytoplankton. Beneficial effects of Al on terrestrial plants (such as tea trees) have been
17 reported, and several mechanisms including increasing Fe availability, promotion of
18 nitrogen, phosphorus and potassium uptake, protection against copper, manganese and
19 phosphorus toxicity, and alleviation of H⁺ toxicity have been proposed for different plant
20 genotypes and growth media (reviews in Foy (1984); Osaki et al. (1997)). As no

1 established biological function of Al has yet been found, the suggestion of indirect roles
2 of Al in the enhanced growth of marine phytoplankton is more readily accepted by most
3 of us who are accustomed to thinking of Al as a non-essential element that is toxic to
4 freshwater organisms and terrestrial plants and animals. However, we suggest that our
5 thinking and exploration of the roles of Al in the enhanced growth of marine
6 phytoplankton should not be constrained by established mechanisms of action, since our
7 understanding of Al effects on marine organisms is still embryonic and limited.

8

9 **5 Possible links of Al to the biological pump and global carbon cycle**

10 More and more evidence shows that Al could play an important role in the biological
11 pump and the global carbon cycle, by influencing marine phytoplankton growth and
12 biogenic matter decay. First, the beneficial effects of Al on marine phytoplankton growth,
13 as described in section 4, indicate that Al could increase carbon fixation in the upper
14 layers of the ocean. On the one hand, by facilitating the uptake of Fe by marine
15 phytoplankton, Al could enhance carbon fixation in the high-nutrient-low-chlorophyll
16 (HNLC) oceans such as the Southern Ocean, and nitrogen fixation and carbon fixation in
17 the oligotrophic tropical and subtropical oceans. On the other hand, by enhancing
18 phytoplankton utilization of DOP, Al could increase carbon fixation in coastal seawaters
19 such as estuaries, where phosphorus limitation can sporadically occur (Lin et al. 2016),
20 and in phosphorus-limited oligotrophic seawater such as the tropical Atlantic Ocean

1 (Browning et al. 2017). The Al-related enhanced carbon fixation in the upper oceans
2 would provide increased organic carbon could potentially be exported to the deep ocean
3 through the biological pump.

4 Second, Al could increase the proportion of organic carbon fixed in the upper ocean
5 that is exported and sequestered in the deep ocean, by reducing the decomposition and
6 decay of particulate biogenic matter. It is well known that Al can reduce the solubility of
7 diatom frustules when it is incorporated into them (Beck et al. 2002; Dixit et al. 2001;
8 Sarmiento and Gruber 2006; Van Cappellen et al. 2002), potentially leading to an
9 increased flux of diatom frustules and associated organic matter into the deep ocean. In
10 addition, Al has been reported to be the most abundant trace metal in fecal pellets
11 produced by zooplankton (e.g., salps) (Cabanes et al. 2017); we suggest that the presence
12 of Al could reduce the solubility and degradation of these pellets, which show higher
13 export efficiencies to the deep ocean than the bulk of sinking particulate organic carbon
14 (Anderson 1998; Cabanes et al. 2017). According to our recent unpublished laboratory
15 data on the microbial degradation of senescent marine diatom cells, the decay of
16 particulate organic carbon fixed by the diatoms was delayed in the presence of Al, and
17 this was accompanied by an increase in cell size and cellular carbon content. All of these
18 effects of Al on particulate biogenic matter would tend to increase the efficiency of the
19 biological pump, leading to increased export of particulate organic carbon produced in
20 the upper ocean to the deep ocean.

1 Third, Al can substitute for other cations (e.g., Mg) in biological systems, reacting
2 with biomolecules to form strong bonds that are slow to dissociate (Exley and Mold 2015;
3 Williams 1996). In addition, Al can induce the formation of strong structures in a wide
4 range of peptides, through binding amino acid sidechains (Mujika et al. 2017). Binding to
5 the peptide backbone has also been considered (Mujika et al. 2014; Song et al. 2014) but
6 is less favored (Mujika et al. 2017). As a result, Al may make organic carbon more
7 difficult to decompose, again leading to greater and longer sequestration in the ocean
8 depths.

9 In summary, we suggest that Al likely plays an unrecognized but important role in the
10 marine and global carbon cycles, not only by enhancing CO₂ fixation in the upper ocean,
11 but also by favouring the export and sequestration of the fixed carbon to the deep ocean.

12

13 **6 The relationship of Al with climate change and Iron Hypothesis**

14 The possible links of Al to the biological pump and the global carbon cycle, and the
15 inverse correlation of Al inputs into the Southern Ocean with atmospheric CO₂ in
16 geological times, indicate that Al may play an important role in climate change.

17

1

2 **6.1 A flaw in the Iron Hypothesis**

3 The Iron Hypothesis was proposed by John Martin, who considered that higher Fe
4 (dust) input into the Southern Ocean was associated with cooler temperatures and lower
5 atmospheric CO₂ concentrations in geological times. This hypothesis was based on the
6 data recorded in the Vostok ice core, which indicated that Fe inputs into the Southern
7 Ocean were inversely correlated with atmospheric CO₂ concentrations over the past 160
8 thousand years (Martin 1990). The hypothesis led to the idea that adding Fe to the ocean
9 surface waters in the Fe-limited HNLC oceans such as the Southern Ocean would
10 stimulate the growth of marine phytoplankton such as diatoms, drawing down
11 atmospheric CO₂ and in turn cooling the planet. More and more paleo-oceanographic
12 data (Abelmann et al. 2006; Kumar et al. 1995; Lambert et al. 2008; Martínez-García et
13 al. 2014; Martínez-García et al. 2011), as well as modern evidence from natural Fe
14 fertilization through dust deposition or upwelling in the Fe-limited HNLC oceans (Bishop
15 et al. 2002; Blain et al. 2007; Han et al. 2006; Pollard et al. 2009), support the proposed
16 link of Fe to climate change, i.e., inputs of Fe into the HNLC oceans would result in
17 phytoplankton blooms in the upper layer, and carbon export and sequestration in the deep
18 ocean.

19 However, the Iron Hypothesis is only partly supported by experimental ocean Fe
20 fertilizations conducted in Fe-limited seawaters all over the world. To examine the idea

1 that Fe fertilization might control past and future climate, as predicted by the Iron
2 Hypothesis, 13 ocean Fe fertilization experiments in Fe-limited seawaters, such as the
3 Southern Ocean, have been completed since 1993. Adding Fe indeed brought about
4 phytoplankton blooms in the upper ocean, but it did not necessarily result in increased
5 carbon export and sequestration into the deep ocean (Boyd et al. 2007; Boyd et al. 2004;
6 Martin et al. 2013). In contrast, enhanced carbon export to the deep ocean was
7 consistently observed following natural Fe fertilization; natural Fe fertilization was 10 to
8 100 times more efficient at removing carbon from the surface ocean than any artificial Fe
9 fertilization has been (Blain et al. 2007; Pollard et al. 2009; Salter et al. 2014; Wolff et al.
10 2011). This observation led us to consider that Fe alone could not adequately explain the
11 drawdown of atmospheric CO₂ concentrations and cooler temperatures in glacial periods,
12 i.e., there could be a flaw in the Iron Hypothesis, and the differences between natural and
13 experimental Fe fertilizations may tell us where the flaw lies.
14

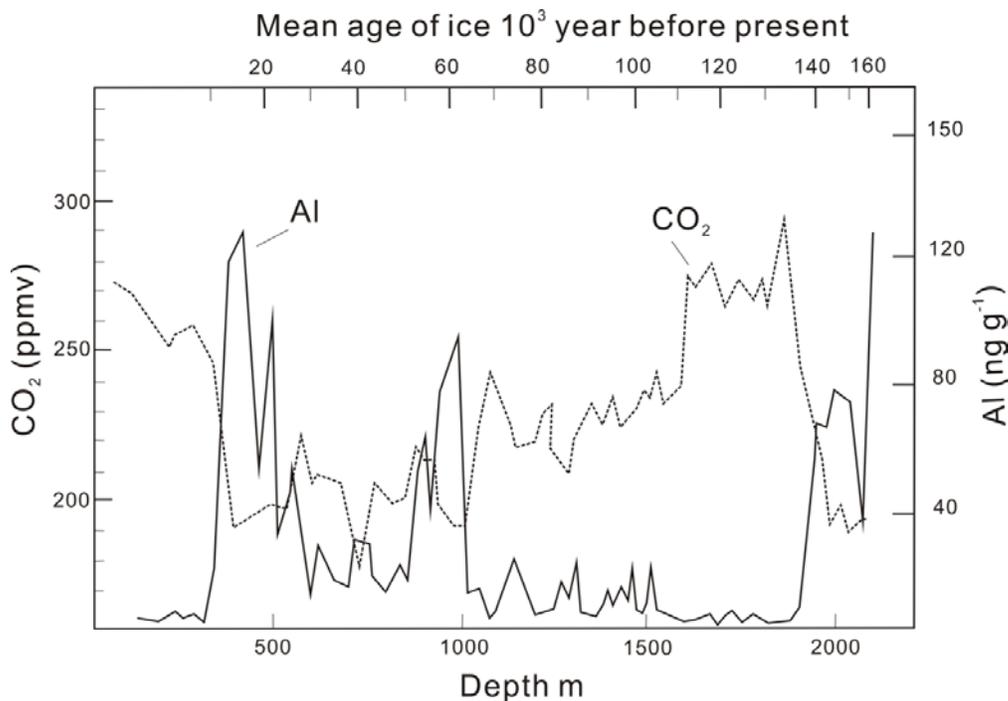
1 **6.2 A revised Iron-Aluminum Hypothesis**

2 Natural Fe fertilization, as caused by dust deposition, upwelling and hydrothermal
3 venting, provides the ocean not only Fe, but also Al and other elements. Silicate and
4 phosphorus from dust deposition have been suggested to favor net primary production
5 and a greater carbon sink in the ocean (Falkowski et al. 1998; Harrison 2000). It is also
6 well established that dust deposition is the major source of dissolved Al in the upper layer
7 of the open ocean, most of this Al being in the form of aluminosilicates. Several studies
8 have reported the solubility of Al in dust aerosols (e.g., Baker et al. 2006; Buck et al.
9 2013; Measures et al. 2010). The median solubility is around 5%, and 80% of the
10 dissolvable Al in dust aerosols is released rapidly in seawater, within the first 24 h, and a
11 further 20% is released slowly over the next three days (Measures et al. 2010). We here
12 propose that the Al associated with natural Fe fertilization may be responsible for the
13 inconsistency between natural and experimental Fe fertilization events.

- 14 • Re-examining the argument for the Iron Hypothesis, we find that the original data
15 from the Vostok ice core showed an inverse correlation not between Fe and CO₂, but
16 rather between Al and CO₂ (Fig. 3). Martin converted the original Al data to Fe
17 estimates using Taylor's (1964) crustal abundance values (Martin 1990). In other
18 words, the original data also support the idea that higher Al inputs into the Southern
19 Ocean were associated with cooler temperatures and lower atmospheric CO₂
20 concentrations in geological times (De Angelis et al. 1987; Petit et al. 1999).

- 1 • As discussed earlier, with reference to the effect of Al on various biogeochemical
- 2 processes (Section 4), this Al could allow marine phytoplankton to use the Fe inputs
- 3 from natural processes like dust deposition or hydrothermal venting more efficiently.
- 4 • Finally, the ability of Al to reduce the decomposition and decay of biogenic matter
- 5 could favor the export of the Fe fertilization-generated organic carbon in HNLC
- 6 oceans to the deep ocean. Moreover, Al could also potentially increase carbon export
- 7 and sequestration in non-HNLC oceans by stimulating nitrogen fixation and
- 8 facilitating the utilization of Fe and DOP by marine phytoplankton, as discussed in
- 9 Section 4.

10



11

12 **Figure 3.** Al concentrations as a function of real depth in the Antarctic Vostok ice core,

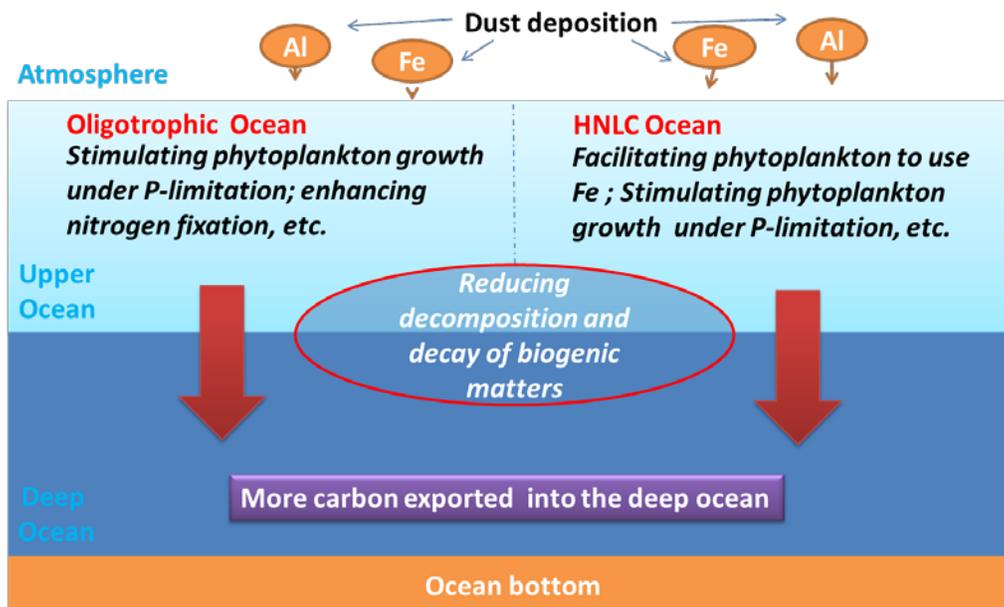
13

together with mean CO₂ concentrations in air trapped in ice, versus mean age

1 of ice. Original figure adapted from Martin (1990). Al data are from De
2 Angelis et al. (1987); CO₂ concentrations are from Barnola et al. (1987).

3

4 Accordingly, we would like to propose a revision to the original Iron Hypothesis, to
5 propose a new Fe-Al hypothesis that takes into account the roles played by Al as well as
6 Fe in the glacial-interglacial change in atmospheric CO₂ concentrations and climate
7 change. In the new hypothesis, we do not deny the important role of Fe, but seek
8 recognition that the role of Al is also important in the global carbon cycle, as outlined in
9 Fig. 4. We predict that adding both Al and Fe in the HNLC oceans such as the Southern
10 Ocean, will result not only in phytoplankton blooms in the upper ocean, but also high
11 carbon export and sequestration to the deep ocean.



12

1 **Figure 4.** Schematic description of the revised Fe-Al Hypothesis. Natural Fe fertilization
2 processes like (but not limited to) dust deposition provide the ocean not only
3 with Fe but also Al; Al, as well as Fe, plays an important role in the
4 glacial-interglacial change in atmospheric CO₂ concentration and climate
5 change.

6
7
8
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1 **References**

- 2 Abelman A, Gersonde R, Cortese G, Kuhn G, Smetacek V (2006) Extensive
3 phytoplankton blooms in the Atlantic sector of the glacial Southern Ocean.
4 *Paleoceanography* 21:PA1013. doi: 10.1029/2005pa001199
- 5 Anderson V (1998) Salp and pyrosomid blooms and their importance in biogeochemical
6 cycles. In: Bone Q (ed) *The Biology of Pelagic Tunicates*. Oxford University
7 Press, Oxford, pp 125-137
- 8 Angel BM, Apte SC, Batley GE, Golding LA (2016) Geochemical controls on aluminium
9 concentrations in coastal waters. *Environ Chem* 13:111-118. doi:
10 10.1071/EN15029
- 11 Baker AR, Jickells TD, Witt M, Linge KL (2006) Trends in the solubility of iron,
12 aluminium, manganese and phosphorus in aerosol collected over the Atlantic
13 Ocean. *Mar Chem* 98: 43-58. doi: 10.1016/j.marchem.2005.06.004
- 14 Barnola JM, Raynaud D, Korotkevich YS, Lorius C (1987) Vostok ice core provides
15 160,000-year record of atmospheric CO₂. *Nature* 329: 408-414. doi:
16 10.1038/329408a0
- 17 Beardmore J, Lopez X, Mujika JI, Exley C (2016) What is the mechanism of formation
18 of hydroxyaluminosilicates? *Sci Rep* 6: 8. doi: 10.1038/srep30913
- 19 Beck L, Gehlen M, Flank AM, Van Bennekom AJ, Van Beusekom JEE (2002) The
20 relationship between Al and Si in biogenic silica as determined by PIXE and XAS.
21 *Nucl Instrum Methods Phys Res Sect B* 189:180-184. doi:
22 10.1016/s0168-583x(01)01035-7
- 23 Bengtsson B-E (1978) Use of a harpacticoid copepod in toxicity tests. *Mar Pollut Bull*
24 9:238-241. doi: 10.1016/0025-326X(78)90378-8

- 1 Bishop JKB, Davis RE, Sherman JT (2002) Robotic observations of dust storm
2 enhancement of carbon biomass in the North Pacific. *Science* 298:817-821. doi:
3 10.1126/science.1074961
- 4 Blain S et al. (2007) Effect of natural iron fertilization on carbon sequestration in the
5 Southern Ocean. *Nature* 446:1070-1074. doi: 10.1038/nature05700
- 6 Boyd PW et al. (2007) Mesoscale iron enrichment experiments 1993-2005: synthesis and
7 future directions. *Science* 315:612-617. doi: 10.1126/science.1131669
- 8 Boyd PW et al. (2004) The decline and fate of an iron-induced subarctic phytoplankton
9 bloom. *Nature*:1-4. doi: 10.1038/nature02437
- 10 Buck CS, Landing WM, Resing J (2013) Pacific Ocean aerosols: Deposition and
11 solubility of iron, aluminum, and other trace elements. *Mar Chem* 157: 117-130.
12 doi: 10.1016/j.marchem.2013.09.005
- 13 Browning TJ, Achterberg EP, Yong JC, Rapp I, Utermann C, Engel A, Moore CM (2017)
14 Iron limitation of microbial phosphorus acquisition in the tropical North Atlantic.
15 *Nature Commun* 8:15465. doi: 10.1038/ncomms15465
- 16 Cabanes DJE, Norman L, Santos-Echeandía J, Iversen MH, Trimborn S, Laglera LM,
17 Hassler CS (2017) First evaluation of the role of salp fecal pellets on iron
18 biogeochemistry. *Front Mar Sci* 3. doi: 10.3389/fmars.2016.00289
- 19 Calabrese A, Collier RS, Nelson DA, MacInnes JR (1973) The toxicity of heavy metals
20 to embryos of the American oyster *Crassostrea virginica*. *Mar Biol* 18:162-166.
21 doi: 10.1007/BF00367984
- 22 Campbell PGC, Hansen HJ, Dubreuil B, Nelson WO (1992) Geochemistry of Quebec
23 North Shore salmon rivers during snowmelt: organic acid pulse and aluminum
24 mobilization. *Can J Fish Aquat Sci* 49:1938-1952. doi: 10.1139/f92-215

- 1 Caplat C, Oral R, Mahaut ML, Mao A, Barillier D, Guida M, Della Rocca C, Pagano G
2 (2010) Comparative toxicities of aluminum and zinc from sacrificial anodes or
3 from sulfate salt in sea urchin embryos and sperm. *Ecotox Environ Saf*
4 73:1138-1143. doi: 10.1016/j.ecoenv.2010.06.024
- 5 Collier R, Edmond J (1984) The trace element geochemistry of marine biogenic
6 particulate matter. *Prog Oceanogr* 13:113-199. doi:
7 10.1016/0079-6611(84)90008-9
- 8 Crémazy A, Campbell PGC, Fortin C (2013) The Biotic Ligand Model can successfully
9 predict the uptake of a trivalent Ion by a unicellular alga below pH 6.50 but not
10 above: possible role of hydroxo-species. *Environ Sci Technol* 47:2408-2415. doi:
11 10.1021/es3038388
- 12 da Cunha LC, Croot P, LaRoche J (2009) Influence of river discharge in the tropical and
13 subtropical North Atlantic Ocean. *Limnol Oceanogr* 54:644-648. doi:
14 10.4319/lo.2009.54.2.0644
- 15 De Angelis M, Barkov N, Petrov V (1987) Aerosol concentrations over the last climatic
16 cycle (160 kyr) from an Antarctic ice core. *Nature* 325:318-321. doi:
17 10.1038/325318a0
- 18 Diaz JM, Hansel CM, Voelker BM, Mendes CM, Andeer PF, Zhang T (2013)
19 Widespread production of extracellular superoxide by heterotrophic bacteria.
20 *Science* 340:1223-1226. doi: 10.1126/science.1237331
- 21 Dixit S, Van Cappellen P, van Bennekom AJ (2001) Processes controlling solubility of
22 biogenic silica and pore water build-up of silicic acid in marine sediments. *Mar*
23 *Chem* 73:333-352. doi: 10.1016/S0304-4203(00)00118-3

- 1 Driscoll CT, Schecher WD (1990) The chemistry of aluminum in the environment
2 Environ Geochem Health 12:28-49. doi: 10.1007/BF01734046
- 3 Ebihara N, Uchida T, Ogura H, Oguma K (2006) Determination of metal elements in
4 marine planktons after pressurized acid decomposition Bunseki Kagaku
5 55:855-861. doi: 10.2116/bunsekikagaku.55.855
- 6 Elderfield H, Schultz A (1996) Mid-ocean ridge hydrothermal fluxes and the chemical
7 composition of the ocean. Ann Rev Earth Planet Sci 24: 191-224. doi:
8 10.1146/annurev.earth.24.1.191
- 9 Exley C (2004) The pro-oxidant activity of aluminum. Free Radical Bio Med 36:380-387.
10 doi: 10.1016/j.freeradbiomed.2003.11.017
- 11 Exley C (2012) Reflections upon and recent insight into the mechanism of formation of
12 hydroxyaluminosilicates and the therapeutic potential of silicic acid. Coord Chem
13 Rev 256: 82-88. doi: 10.1016/j.ccr.2011.06.022
- 14 Exley C, Mold MJ (2015) The binding, transport and fate of aluminium in biological cells.
15 J Trace Elem Med Bio 30:90-95. doi: 10.1016/j.jtemb.2014.11.002
- 16 Falkowski PG, Barber RT, Smetacek V (1998) Biogeochemical controls and feedbacks
17 on ocean primary production. Science 281: 200-206. doi:
18 10.1126/science.281.5374.200
- 19 Foy CD (1984) Physiological effects of hydrogen, aluminum, and manganese toxicities in
20 acid soil. In: Adams F (ed) Soil Acidity and Liming, 2nd edn. pp 57-97. American
21 Society of Agronomy, Madison, WI, USA
- 22 Gehlen M, Beck L, Calas G, Flank AM, Van Bennekom AJ, Van Beusekom JEE (2002)
23 Unraveling the atomic structure of biogenic silica: evidence of the structural

1 association of Al and Si in diatom frustules. *Geochim Cosmochim Acta*
2 66:1601-1609. doi: 10.1016/s0016-7037(01)00877-8

3 Gensemer RW, Playle RC (1999) The bioavailability and toxicity of aluminum in aquatic
4 environments. *Crit Rev Environ Sci Technol* 29:315-450. doi:
5 10.1080/10643389991259245

6 Gillmore ML, Golding LA, Angel BM, Adams MS, Jolley DF (2016) Toxicity of
7 dissolved and precipitated aluminium to marine diatoms. *Aquat Toxicol*
8 174:82-91. doi: 10.1016/j.aquatox.2016.02.004

9 Golding LA, Angel BM, Batley GE, Apte SC, Krassoi R, Doyle CJ (2015) Derivation of
10 a water quality guideline for aluminium in marine waters. *Environ Toxicol Chem*
11 34:141-151. doi: 10.1002/etc.2771

12 Hall IR, Hydes DJ, Statham PJ, Overnell J (1999) Seasonal variations in the cycling of
13 aluminium, cadmium and manganese in a Scottish sea loch: biogeochemical
14 processes involving suspended particles. *Cont Shelf Res* 19:1783-1808. doi:
15 10.1016/S0278-4343(99)00056-4

16 Han Q, Moore JK, Zender C, Measures C, Hydes D (2008) Constraining oceanic dust
17 deposition using surface ocean dissolved Al. *Global Biogeochem Cycles*
18 22:GB2003. doi: 10.1029/2007gb002975

19 Han Y, Fang X, Xi X, Song L, Yang S (2006) Dust storm in Asia continent and its
20 bio-environmental effects in the North Pacific: A case study of the strongest dust
21 event in April, 2001 in central Asia. *Chin Sci Bull* 51:723-730. doi:
22 10.1007/s11434-006-0723-2

- 1 Hansel CM, Buchwald C, Ossolinski J, Dyhrman S, Van Mooy B, Polyviou D (2016)
2 Dynamics of extracellular superoxide production by *Trichodesmium* colonies
3 from the Sargasso Sea. *Limnol Oceanogr* 61:1188-1200. doi: 10.1002/lno.10266
- 4 Harford AJ, Hogan AC, Tsang JJ, Parry DL, Negri AP, Adams MS, Stauber JL, van Dam
5 RA (2011) Effects of alumina refinery wastewater and signature metal
6 constituents at the upper thermal tolerance of: 1. The tropical diatom *Nitzschia*
7 *closterium* *Mar Pollut Bull* 62:466-473. doi: 10.1016/j.marpolbul.2011.01.013
- 8 Harrison KG (2000) Role of increased marine silica input on paleo- $p\text{CO}_2$ levels.
9 *Paleoceanography* 15: 292-298. doi: 10.1029/1999PA000427
- 10 Kaupp LJ, Measures CI, Selph KE, Mackenzie FT (2011) The distribution of dissolved
11 Fe and Al in the upper waters of the Eastern Equatorial Pacific. *Deep-Sea Res II*:
12 58:296-310. doi: 10.1016/j.dsr2.2010.08.009
- 13 Khan A, Dobson JP, Exley C (2006) Redox cycling of iron by *A beta*(42). *Free Radical*
14 *Bio Med* 40: 557-569. doi: 10.1016/j.freeradbiomed.2005.09.013
- 15 Kondo Y, Moffett JW (2015) Iron redox cycling and subsurface offshore transport in the
16 eastern tropical South Pacific oxygen minimum zone *Mar Chem* 168:95-103. doi:
17 10.1016/j.marchem.2014.11.007
- 18 Koning E, Gehlen M, Flank AM, Calas G, Epping E (2007) Rapid post-mortem
19 incorporation of aluminum in diatom frustules: evidence from chemical and
20 structural analyses. *Mar Chem* 106:208-222. doi: 10.1016/j.marchem.2006.06.009
- 21 Kumar N, Anderson RF, Mortlock RA, Froelich PN, Kubik P, Dittrich-Hannen B, Suter
22 M (1995) Increased biological productivity and export production in the glacial
23 Southern Ocean. *Nature* 378:675-680. doi: 10.1038/378675a0

- 1 Kuss J, Kremling K (1999) Spatial variability of particle associated trace elements in
2 near-surface waters of the North Atlantic (30°N/60°W to 60°N/2°W), derived by
3 large volume sampling. *Mar Chem* 68:71-86. doi:
4 10.1016/S0304-4203(99)00066-3
- 5 Lambert F , Delmonte B, Petit JR, Bigler M, Kaufmann PR, Hutterli MA, Stocker TF,
6 Ruth U, Steffensen JP, Maggi V (2008) Dust-climate couplings over the past
7 800,000 years from the EPICA Dome C ice core. *Nature* 452:616-619. doi:
8 10.1038/nature06763
- 9 Leleyter L, Baraud F, Gil O, Gouali S, Lemoine M, Orvain F (2016) Al impact on growth
10 of benthic diatoms. In: Williams S (ed) *Marine Sediments: Formation,*
11 *Distribution and Environmental Impacts*, Nova Science Publishers, pp 61-79
- 12 Lin S, Litaker RW, Sunda WG (2016) Phosphorus physiological ecology and molecular
13 mechanisms in marine phytoplankton. *J Phycol* 52:10-36. doi: 10.1111/jpy.12365
- 14 Liu J, Zhou L, Ke Z, Li G, Shi R, Tan Y (2017a) Effect of aluminum on the growth and
15 nitrogen fixation of a marine nitrogen-fixing cyanobacterium: *Crocospaera*
16 *watsonii* (In Chinese with English abstract). *J Trop Oceanogr* 36:12-18. doi:
17 10.11978/2016069
- 18 Liu J, Zhou L, Li G, Ke Z, Shi R, Tan Y (2018) Beneficial effects of aluminum
19 enrichment on nitrogen-fixing cyanobacteria in the South China Sea. *Mar Pollut*
20 *Bull* In press. doi: 10.1016/j.marpolbul.2018.02.011
- 21 Liu J, Zhou L, Tan Y, Wang Q, Hu Z, Li J, Jiang X, Ke Z (2017b) Distribution of
22 reactive aluminum under the influence of mesoscale eddies in the western South
23 China Sea. *Acta Oceanol Sin* 36:95-103. doi: 10.1007/s1313

- 1 Macdonald JM, Shields JD, Zimmer-Faust RK (1988) Acute toxicities of eleven metals to
2 early life-history stages of the yellow crab *Cancer anthonyi*. Mar Biol 98:201-207.
3 doi: 10.1007/BF00391195
- 4 Macdonald TL, Martin RB (1988) Aluminum ion in biological systems. Trends Biochem
5 Sci 13:15-19. doi: 10.1016/0968-0004(88)90012-6
- 6 Machill S, Kohler L, Ueberlein S, Hedrich R, Kunaschk M, Paasch S, Schulze R,
7 Brunner E (2013) Analytical studies on the incorporation of aluminium in the cell
8 walls of the marine diatom *Stephanopyxis turris*. BioMetals 26:141-150. doi:
9 10.1007/s10534-012-9601-3
- 10 Maring HB, Duce RA (1987) The impact of atmospheric aerosols on trace metal
11 chemistry in open ocean surface seawater, 1. Aluminum. Earth Planet Sci Lett
12 84:381-392. doi: 10.1016/0012-821X(87)90003-3
- 13 Martínez-García A, Sigman DM, Ren HJ, Anderson RF, Straub M, Hodell DA, Jaccard
14 SL, Eglinton TI, Haug GH (2014) Iron fertilization of the subantarctic Ocean
15 during the Last Ice Age. Science 343:1347-1350. doi: 10.1126/science.1246848
- 16 Martínez-García A, Rosell-Melé A, Jaccard SL, Geibert W, Sigman DM, Haug GH (2011)
17 Southern Ocean dust-climate coupling over the past four million years. Nature
18 476:312-315. doi: 10.1038/nature10310
- 19 Martin JH (1990) Glacial-interglacial CO₂ change: the Iron Hypothesis.
20 Paleoceanography 5:1-13. doi: 10.1029/PA005i001p00001
- 21 Martin JH, Knauer GA (1973) The elemental composition of plankton. Geochim
22 Cosmochim Acta 37:1639-1653. doi: 10.1016/0016-7037(73)90154-3

- 1 Martin P et al. (2013) Iron fertilization enhanced net community production but not
2 downward particle flux during the Southern Ocean iron fertilization experiment
3 LOHAFEX. *Global Biogeochem Cycles* 27:871-881. doi: 10.1002/gbc.20077
- 4 Measures CI, Sato T, Vink S, Howell S, Li YH (2010) The fractional solubility of
5 aluminium from mineral aerosols collected in Hawaii and implications for
6 atmospheric deposition of biogeochemically important trace elements. *Marine*
7 *Chem* 120: 144-153. doi: 10.1016/j.marchem.2009.01.014
- 8 Measures C, Hatta M, Fitzsimmons J, Morton P (2015) Dissolved Al in the zonal N
9 Atlantic section of the US GEOTRACES 2010/2011 cruises and the importance
10 of hydrothermal inputs. *Deep-Sea Res II* 116:176-186. doi:
11 10.1016/j.dsr2.2014.07.006
- 12 Menzel DW, Hulburt EM, Tyther JH (1963) The effects of enriching Sargasso sea water
13 on the production and species composition of the phytoplankton. *Deep Sea Res*
14 *Oceanogr Abstr* 10:209-219. doi: 10.1016/0011-7471(63)90357-7
- 15 Middag R, van Slooten C, de Baar HJW, Laan P (2011) Dissolved aluminium in the
16 Southern Ocean. *Deep-Sea Res II* 58:2647-2660. doi: 10.1016/j.dsr2.2011.03.001
- 17 Millero FJ (2009) Effect of ocean acidification on the speciation of metals in seawater.
18 *Oceanography* 22: 72-85. doi: 10.5670/oceanog.2009.98
- 19 Moffett JW, Goepfert TJ, Naqvi SWA (2007) Reduced iron associated with secondary
20 nitrite maxima in the Arabian Sea. *Deep-Sea Res I* 54:1341-1349. doi:
21 10.1016/j.dsr.2007.04.004
- 22 Moore CM et al. (2009) Large-scale distribution of Atlantic nitrogen fixation controlled
23 by iron availability. *Nature Geosci* 2:867-871. doi: 10.1038/NGEO667

- 1 Moran SB, Moore RM (1988) Evidence from mesocosm studies for biological removal of
2 dissolved aluminium from sea water. *Nature* 335:706-708. doi: 10.1038/335706a0
- 3 Mujika JI, Dalla Torre G, Formoso E, Grande-Aztatzi R, Grabowski SJ, Exley C, Lopez
4 X (2017) Aluminum's preferential binding site in proteins: sidechain of amino
5 acids versus backbone interactions. *J Inorg Biochem.* doi:
6 10.1016/j.jinorgbio.2017.10.014
- 7 Mujika JI, Rezabal E, Mercero JM, Ruipérez F, Costa D, Ugalde JM, Lopez X (2014)
8 Aluminium in biological environments: a computational approach. *Computat*
9 *Struct Biotechnol J* 9:e201403002. doi: 10.5936/csbj.201403002
- 10 Mujika JI, Ruipérez F, Infante I, Ugalde JM, Exley C, Lopez X (2011) Pro-oxidant
11 activity of aluminum: stabilization of the aluminum superoxide radical ion. *J Phys*
12 *Chem A* 115:6717-6723. doi: 10.1021/jp203290b
- 13 Negri AP, Harford AJ, Parry DL, van Dam RA (2011) Effects of alumina refinery
14 wastewater and signature metal constituents at the upper thermal tolerance of: 2.
15 The early life stages of the coral *Acropora tenuis*. *Mar Pollut Bull* 62:474-482.
16 doi: 10.1016/j.marpolbul.2011.01.011
- 17 Orians KJ, Bruland KW (1986) The biogeochemistry of aluminum in the Pacific Ocean.
18 *Earth Planet Sci Lett* 78: 397-410. doi: 10.1016/0012-821X(86)90006-3
- 19 Osaki M, Watanabe T, Tadano T (1997) Beneficial effect of aluminum on growth of
20 plants adapted to low pH soils. *Soil Sci Plant Nutr* 43:551-563. doi:
21 10.1080/00380768.1997.10414782
- 22 Pagano G, His E, Beiras R, DeBiase A, Korkina LG, Iaccarino M, Oral R, Quiniou F,
23 Warnau M. Trieff NM (1996) Cytogenetic, developmental, and biochemical

- 1 effects of aluminum, iron, and their mixture in sea urchins and mussels. Arch
2 Environ Contam Toxicol 31:466-474. doi: 10.1007/BF00212429
- 3 Petit J-R et al. (1999) Climate and atmospheric history of the past 420,000 years from the
4 Vostok ice core, Antarctica. Nature 399:429-436. doi: 10.1038/20859
- 5 Petrich SM, Reish DJ (1979) Effects of aluminium and nickel on survival and
6 reproduction in polychaetous annelids. Bull Environ Contam Toxicol 23:698-702.
7 doi: 10.1007/BF01770027
- 8 Pierrot D, Millero FJ (2017) The speciation of metals in natural waters. Aquat Geochem
9 23:1-20. doi: 10.1007/s10498-016-9292-4
- 10 Planquette H, Fones GR, Statham PJ, Morris PJ (2009) Origin of iron and aluminium in
11 large particles (>53 µm) in the Crozet region, Southern Ocean. Mar Chem
12 115:31-42. doi: 10.1016/j.marchem.2009.06.002
- 13 Pollard RT et al. (2009) Southern Ocean deep-water carbon export enhanced by natural
14 iron fertilization. Nature 457:577-580. doi: 10.1038/nature07716
- 15 Ren J-L, Zhang GL, Zhang J, Shi JH, Liu SM, Li FM, Jin J, Liu CG (2011) Distribution
16 of dissolved aluminum in the Southern Yellow Sea: influences of a dust storm and
17 the spring bloom. Mar Chem 125:69-81. doi: 10.1016/j.marchem.2011.02.004
- 18 Rengel Z (1996) Uptake of aluminium by plant cells. New Phytologist 134:389-406. doi:
19 10.1111/j.1469-8137.1996.tb04356.x
- 20 Resing JA, Sedwick PN, German CR, Jenkins WJ, Moffett JW, Sohst BM, Tagliabue A
21 (2015) Basin-scale transport of hydrothermal dissolved metals across the South
22 Pacific Ocean. Nature 523:200-203. doi: 10.1038/nature14577

- 1 Rijkenberg MJA, Langlois RJ, Mills MM, Patey MD, Hill PG, Nielsdottir MC, Compton
2 TJ, LaRoche J, Achterberg EP (2011) Environmental forcing of nitrogen fixation
3 in the eastern tropical and sub-tropical North Atlantic Ocean PLoS ONE 6:e28989.
4 doi: 10.1371/journal.pone.0028989
- 5 Rose AL (2012) The influence of extracellular superoxide on iron redox chemistry and
6 bioavailability to aquatic microorganisms. Front Microbiol 3:1-21. doi:
7 10.3389/fmicb.2012.00124
- 8 Rose AL, Waite TD (2005) Reduction of organically complexed ferric iron by superoxide
9 in a simulated natural water. Environ Sci Technol 39:2645-2650. doi:
10 10.1021/es048765k
- 11 Ruipérez F, Mujika JI, Ugalde JM, Exley C, Lopez X (2012) Pro-oxidant activity of
12 aluminum: promoting the Fenton reaction by reducing Fe(III) to Fe(II). J Inorg
13 Biochem 117:118-123. doi: 10.1016/j.jinorgbio.2012.09.008
- 14 Saçan MT, Oztay F, Bolkent S (2007) Exposure of *Dunaliella tertiolecta* to lead and
15 aluminum: Toxicity and effects on ultrastructure. Biol Trace Elem Res
16 120:264-272. doi: 10.1007/s12011-007-8016-4
- 17 Salter I, Schiebel R, Ziveri P, Movellan A, Lampitt R, Wolff GA (2014) Carbonate
18 counter pump stimulated by natural iron fertilization in the Polar Frontal Zone.
19 Nature Geosci 7:885-889. doi: 10.1038/ngeo2285
- 20 Santana-Casiano JM, Gonzalez-Davila M, Laglera LM, Perez-Pena J, Brand L, Millero
21 FJ (1997) The influence of zinc, aluminum and cadmium on the uptake kinetics of
22 iron by algae. Mar Chem 59:95-111. doi: 10.1016/s0304-4203(97)00068-6

- 1 Santos MA (2008) Recent developments on 3-hydroxy-4-pyridinones with respect to
2 their clinical applications: mono and combined ligand approaches. *Coord Chem*
3 *Rev* 252:1213-1224. doi: 10.1016/j.ccr.2008.01.033
- 4 Sarmiento JL, Gruber N (eds) (2006) *Ocean Biogeochemical Dynamics*. Princeton
5 University Press, Princeton, NJ, USA
- 6 Savenko AV, Savenko VS (2011) Aluminum hydroxide's solubility and the forms of
7 dissolved aluminum's occurrence in seawater. *Oceanology* 51:231-234. doi:
8 10.1134/s0001437011020147
- 9 Schneider RJ, Roe KL, Hansel CM, Voelker BM (2016) Species-level variability in
10 extracellular production rates of reactive oxygen species by diatoms. *Front Chem*
11 4:5. doi: 10.3389/fchem.2016.00005
- 12 Shaked Y, Kustka AB, Morel FM (2005) A general kinetic model for iron acquisition by
13 eukaryotic phytoplankton. *Limnol Oceanogr* 50:872-882. doi:
14 10.4319/lo.2005.50.3.0872
- 15 Shi R, Li G, Zhou L, Tan Y (2015) Increasing aluminum alters the growth, cellular
16 chlorophyll a and oxidation stress of cyanobacteria *Synechococcus* sp. *Oceanol*
17 *Hydrobiol Stud* 44:343-351. doi: 10.1515/ohs-2015-0033
- 18 Shi R, Li Z, Zhou L, Tan Y (2016) Influence of dissolved aluminum on marine
19 phytoplankton community structure and growth of *Synechococcus* sp. (In Chinese
20 with English abstract) *South China Fish Sci* 12:1-8. doi:
21 10.3969/j.issn.2095-0780.2016.01.001
- 22 Song B et al. (2014) Irreversible denaturation of proteins through aluminum-induced
23 formation of backbone ring structures. *Angew Chem Int Edit* 53:6358-6363. doi:
24 10.1002/anie.201307955

- 1 Stoffyn M (1979) Biological control of dissolved aluminum in seawater: experimental
2 evidence. *Science* 203:651-653. doi: 10.1126/science.203.4381.651
- 3 Taylor GJ, McDonald-Stephens JL, Hunter DB, Bertsch PM, Elmore D, Rengel Z, Reid
4 RJ (2000) Direct measurement of aluminum uptake and distribution in single cells
5 of *Chara corallina*. *Plant Physiol* 123:987-996. doi: 10.1104/pp.123.3.987
- 6 Taylor SR (1964) Abundance of chemical elements in the continental crust: a new table.
7 *Geochim Cosmochim Acta* 28:1273-1285. doi: 10.1016/0016-7037(64)90129-2
- 8 Teien H-C, Standring WJF, Salbu B (2006) Mobilization of river transported colloidal
9 aluminium upon mixing with seawater and subsequent deposition in fish gills. *Sci*
10 *Total Environ* 364:149-164. doi: 10.1016/j.scitotenv.2006.01.005
- 11 Tovar-Sanchez A, Sanudo-Wilhelmy SA, Kustka AB, Agusti S, Dachs J, Hutchins DA,
12 Capone DG, Duarte CM (2006) Effects of dust deposition and river discharges on
13 trace metal composition of *Trichodesmium* spp. in the tropical and subtropical
14 North Atlantic Ocean. *Limnol Oceanogr* 51:1755-1761. doi:
15 10.4319/lo.2006.51.4.1755
- 16 Trenfield MA, van Dam JW, Harford AJ, Parry D, Streten C, Gibb K, van Dam RA
17 (2015) Aluminium, gallium, and molybdenum toxicity to the tropical marine
18 microalga *Isochrysis galbana*. *Environ Toxicol Chem* 34:1833-1840. doi:
19 10.1002/etc.2996
- 20 Van Bennekom A, Buma A, Nolting R (1991) Dissolved aluminium in the
21 Weddell-Scotia Confluence and effect of Al on the dissolution kinetics of
22 biogenic silica. *Mar Chem* 35:423-434. doi: 10.1016/S0304-4203(09)90034-2

- 1 Van Cappellen P, Dixit S, van Beusekom J (2002) Biogenic silica dissolution in the
2 oceans: reconciling experimental and field-based dissolution rates. *Global*
3 *Biogeochem Cycles* 16:23-21-23-10. doi: 10.1029/2001GB001431
- 4 van Hulst MMP, Sterl A, Tagliabue A, Dutay JC, Gehlen M, de Baar HJW, Middag R
5 (2013) Aluminium in an ocean general circulation model compared with the West
6 Atlantic Geotraces cruises. *J Mar Syst* 126:3-23. doi:
7 10.1016/j.jmarsys.2012.05.005
- 8 Vrieling EG, Poort L, Beelen TPM, Gieskes WWC (1999) Growth and silica content of
9 the diatoms *Thalassiosira weissflogii* and *Navicula salinarum* at different
10 salinities and enrichments with aluminium. *Eur J Phycol* 34:307-316. doi:
11 10.1080/09670269910001736362
- 12 Wang Z, Ren J, Yan L, Xie F, Su R (2013) Preliminary study on scavenging mechanism
13 of dissolved aluminum by phytoplankton. *Acta Ecol Sin* 33:7140-7147. doi:
14 10.5846/stxb201207251057
- 15 Willey JD (1975) Reactions which remove dissolved alumina from seawater. *Mar Chem*
16 3:227-240. doi: 10.1016/0304-4203(75)90004-3
- 17 Williams RJP (1996) Aluminium and biological systems: an introduction. *Coordin Chem*
18 *Rev* 149:1-9. doi: 10.1016/S0010-8545(96)90004-1
- 19 Wilson SP, Hyne RV (1997) Toxicity of acid-sulfate soil leachate and aluminum to
20 embryos of the Sydney rock oyster. *Ecotox Environ Saf* 37:30-36. doi:
21 10.1006/eesa.1996.1514
- 22 Wolff GA et al. (2011) The effects of natural iron fertilisation on deep-sea ecology: the
23 Crozet Plateau, southern Indian Ocean. *PLoS ONE* 6:e20697. doi:
24 10.1371/journal.pone.0020697

- 1 Xie J, Bai XC, Lavoie M, Lu HP, Fan XJ, Pan XL, Fu ZW, Qian HF (2015) Analysis of
2 the proteome of the marine diatom *Phaeodactylum tricornutum* exposed to
3 aluminum providing insights into aluminum toxicity mechanisms Environ Sci
4 Technol 49:11182-11190. doi: 10.1021/acs.est.5b03272
- 5 Yokel RA (2000) The toxicology of aluminum in the brain: a review. Neurotoxicology
6 21:813-828
- 7 Zhang J, Xu H, Yu ZG, Wu Y, Li JF (1999) Dissolved aluminum in four Chinese
8 estuaries: evidence of biogeochemical uncoupling of Al with nutrients. J Asian
9 Earth Sci 17:333-343. doi: 10.1016/S0743-9547(98)00068-3
- 10 Zhou L, Liu J, Tan SXY, Huang L (2017) Phytoplankton responses to aluminum
11 enrichment in the South China Sea. J Inorg Biochem. In press. doi:
12 10.1016/j.jinorgbio.2017.09.022
- 13 Zhou L, Tan Y, Huang L, Wang W-X (2016) Enhanced utilization of organic phosphorus
14 in a marine diatom *Thalassiosira weissflogii*: A possible mechanism for
15 aluminum effect under P limitation. J Exp Mar Biol Ecol 478:77-85. doi:
16 10.1016/j.jembe.2016.02.009