

Table S4. Bacterial strains, plasmids, and primers used in this study.

Bacterial strain or plasmid	Description	Reference or source
<u><i>P. aeruginosa</i> strains</u>		
PAO1	Wild type strain	(1)
PAO1 $\Delta lasR$	PAO1 containing an unmarked, in-frame <i>lasR</i> deletion	(2)
PAO1 $\Delta lasR \Delta rhIR \Delta qscR$	<i>lasR</i> , <i>rhIR</i> , <i>qscR</i> triple mutant in PAO1	(3)
<u>Other strains</u>		
<i>E. coli</i> DH5 α	F ⁻ $\Phi 80 lacZ \Delta M15 \Delta(lacZYA-argF)$ U169 <i>recA1 endA1 hsdR17</i> (r _k ⁻ , m _k ⁺) <i>phoA supE44 thi-1 gyrA96 relA1 λ⁻</i>	Invitrogen
<u>Plasmids</u>		
pSC11	Broad-host-range <i>lasI-lacZ</i> reporter, Ap ^r	(4)
pJN105L	Arabinose inducible <i>lasR</i> in pJN105, Gm ^r	(5)
pECP61.5	Contains an <i>rhIA-lacZ</i> translation fusion and IPTG-inducible <i>rhIR</i> , Ap ^r	(6)
pPROBE-GT	Broad-host-range pVS1/p15a GFP reporter, Gm ^r	(7)
pJF01	pPROBE-GT with -1 through-501 5' region of <i>rhIA</i> inserted with HindIII and BamHI, Gm ^r	This work
pBS351	pPROBE-GT with -1 through-501 5' region of <i>lasI</i> inserted with HindIII and BamHI, Gm ^r	This work
<u>Primers</u>		
<i>lasR</i> .cloning.EcoR1 F	5' – TTTTTGAATTCTAGCGCTATGGCCTTGGTT – 3'	
<i>lasR</i> .cloning.Xba1 R	5' – AAAAATCTAGAGCAAGATCAGAGAGTAATAAGACCC – 3'	
<i>rhIA</i> .GFPcloning.HindIII F	5' – TTTTAAAGCTTGCATGCGAGGCCTGCGAA – 3'	
<i>rhIA</i> .GFPcloning.BamHI R	5' – AAAAAAGGATCCTCTAGAAACCGATACCAACAGACTTTTCGC – 3'	
GFP.QC R	5' – TTCTTTCCTGTACATAACCTTCGGGCA – 3'	

REFERENCES

1. **Stover CK, Pham XQ, Erwin AL, Mizoguchi SD, Warren P, Hickey MJ, Brinkman FS, Hufnagle WO, Kowalik DJ, Lagrou M, Garber RL, Goltry L, Tolentino E, Westbrook-Wadman S, Yuan Y, Brody LL, Coulter SN, Folger KR, Kas A, Larbig K, Lim R, Smith K, Spencer D, Wong GK, Wu Z, Paulsen IT, Reizer J, Saier MH, Hancock RE, Lory S, Olson MV.** 2000. Complete genome sequence of *Pseudomonas aeruginosa* PAO1, an opportunistic pathogen. *Nature* **406**:959-964.
2. **Wang M, Schaefer AL, Dandekar AA, Greenberg EP.** 2015. Quorum sensing and policing of *Pseudomonas aeruginosa* social cheaters. *Proc Natl Acad Sci USA* **112**:2187-2191.
3. **Chugani S, Greenberg EP.** 2010. LuxR homolog-independent gene regulation by acyl-homoserine lactones in *Pseudomonas aeruginosa*. *Proc Natl Acad Sci USA* **107**:10673-10678.
4. **Chugani SA, Whiteley M, Lee KM, D'Argenio D, Manoil C, Greenberg EP.** 2001. QscR, a modulator of quorum-sensing signal synthesis and virulence in *Pseudomonas aeruginosa*. *Proc Natl Acad Sci USA* **98**:2752-2757.
5. **Lee JH, Lequette Y, Greenberg EP.** 2006. Activity of purified QscR, a *Pseudomonas aeruginosa* orphan quorum-sensing transcription factor. *Mol Microbiol* **59**:602-609.
6. **Pearson JP, Pesci EC, Iglewski BH.** 1997. Roles of *Pseudomonas aeruginosa las* and *rhl* quorum-sensing systems in control of elastase and rhamnolipid biosynthesis genes. *J Bacteriol* **179**:5756-5767.
7. **Miller WG, Leveau JH, Lindow SE.** 2000. Improved *gfp* and *inaZ* broad-host-range promoter-probe vectors. *Mol Plant Microbe Interact* **13**:1243-1250.